

CURRENT RESEARCH

ROBERT WEINREB AND EDWARD COTLIER, EDITORS

The Role of the Immune System in Conjunctival Wound Healing After Glaucoma Surgery

L. Chang, FRCOphth,^{1,2} J.G. Crowston, FRCOphth,¹

M. Francesca Cordeiro, PhD, MRCP, FRCOphth,¹

A.N. Akbar, MRCPPath, PhD, BSc,² and P.T. Khaw, PhD, FRCP, FRCS, FRCOphth^{1,3}

¹Wound Healing Research and Glaucoma Units, Institute of Ophthalmology, ²Department of Immunology, Royal Free & University College School of Medicine, ³Department of Pathology and Glaucoma, Moorfields Eye Hospital NHS Trust and Institute of Ophthalmology, London, United Kingdom

Abstract. The immune system has a fundamental role in the development and regulation of ocular healing, which plays an important role in the pathogenesis of most blinding diseases. This review discusses the mechanisms of normal wound healing, describing the animal and fetal wound healing models used to provide further insight into normal wound repair. In particular, conjunctival wound repair after glaucoma filtration surgery will be used to illustrate the contributions that the different components of the immune system make to the healing process. The potential role of macrophages, the possible regulatory effect of lymphocytes, and the important role of growth factors and cytokines in the wound healing reaction are discussed. The significance of the immune system in the pathogenesis of aggressive conjunctival scarring is addressed, particularly assessing the predisposing factors, including drugs, age, and ethnicity. The rationale behind the pharmacological agents currently used to modulate the wound healing response and the effects these drugs have on the function of the immune system are described. Finally, potential new therapeutic approaches to regulating the wound healing response are reported. (Surv Ophthalmol 45:49–68, 2000. © 2000 by Elsevier Science Inc. All rights reserved.)

Key words. anti-metabolites • conjunctival wound healing • corticosteroids • fibroblasts • glaucoma surgery • immune system • macrophages • T lymphocytes

In discussions of conjunctival wound healing, the fibroblast has received the most attention. The purpose of this review is to describe the fundamental and complex role of the immune system in normal conjunctival wound repair and to indicate how it may contribute to excessive wound healing.

The modulation of abnormal conjunctival wound

healing is a subject of great interest, because excessive subconjunctival scarring is the main reason for the failure of glaucoma filtration surgery.² As the successful surgical treatment of glaucoma is probably the most effective way to preserve vision, and indeed, in many parts of the world is often the only practical treatment available, the ability to control the wound healing re-

sponse to maximize the success rate of glaucoma filtration surgery is vitally important.^{130,164} In addition, it is important to understand why some filtration operations fail despite the use of antifibrotic agents.

I. Wound Healing Repair Models

A. ANIMAL MODELS

Healthy conjunctiva is normally populated with cells belonging to the immune system.^{9,56,208} T lymphocytes, macrophages, Langerhans' cells, and occasional B cells have been identified in the epithelium and substantia propria of conjunctival biopsies taken from patients without any primary conjunctival disease. The predominant lymphocyte is the T lymphocyte, with an estimated normal CD4:CD8 T cell ratio of approximately 1:2. During wound healing, there is a rapid and significant increase in inflammatory cell numbers.¹⁹⁹

Various animal models have been used to study conjunctival scarring after glaucoma filtration surgery. We have learned from this work that the immune system must play an important role during the early stages of wound healing.^{166,199} These models show the influx of inflammatory cells, including polymorphonuclear leukocytes, lymphocytes, and macrophages, into the injury site during the first few days following wounding (Fig. 1).

B. FETAL WOUND REPAIR

Further evidence that the immune system participates in wound healing comes from research into fetal wound repair.¹⁶¹ Usually, during the first two trimesters of fetal development, fetal wound healing is not associated with an acute inflammatory reaction, and healing occurs without scar tissue formation.^{5,117} Hopkinson-Woolley et al were not able to demonstrate a macrophage infiltrate in embryonic mouse excisional wounds.¹²⁴ However, after gestational day 14, macrophage recruitment did occur and this was coincident with the stage when fetal scarring began to develop. Interestingly, the addition of cytokines produced by macrophages, such as TGF β or PDGF, to fetal wounds induces an inflammatory response and the development of scar tissue.^{4,145}

II. The Inflammatory Phase of Wound Healing

A. ACTIVATION OF THE IMMUNE SYSTEM IN WOUND HEALING

The wound healing response can be subdivided into a sequence of specific events, made up of three stages: the inflammatory stage, the proliferative stage, and the maturation stage. The immune system

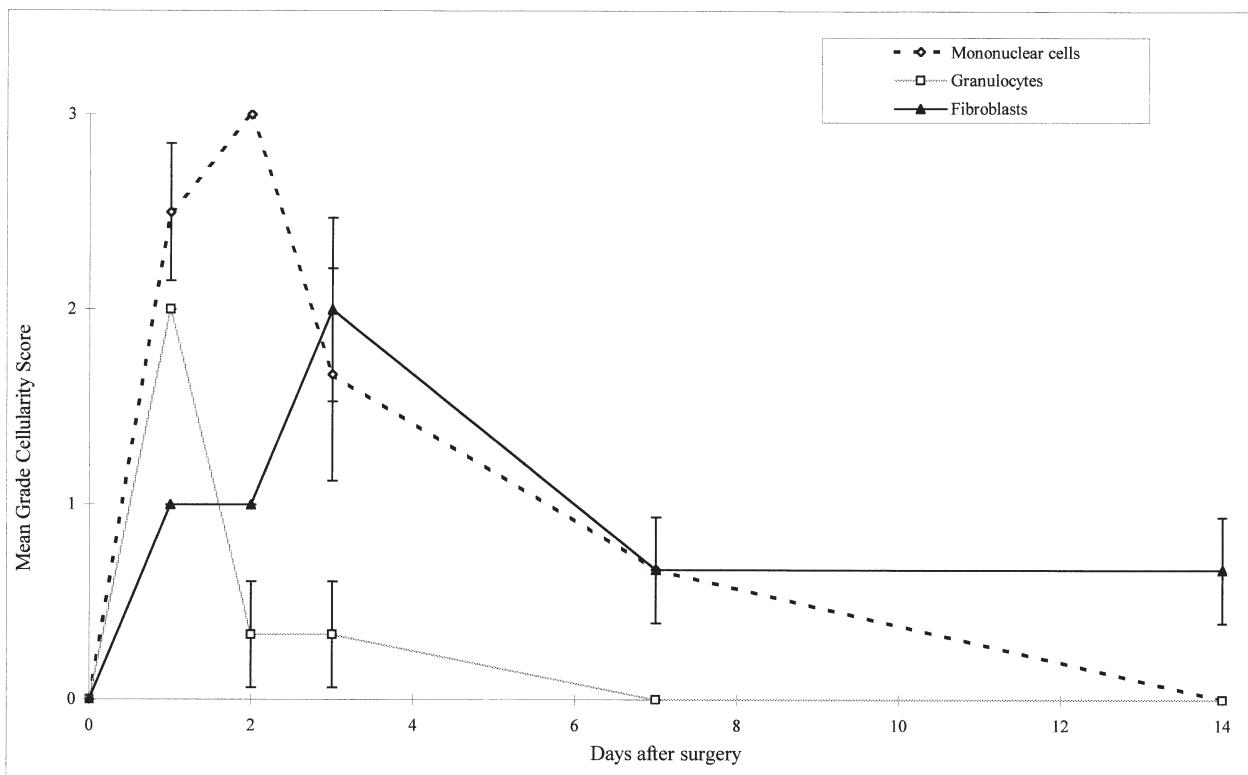


Fig. 1. The cellular profile of mononuclear cells, granulocytes and fibroblast activity, with consecutive peaks consistent with the classic wound healing response in a mouse model of conjunctival scarring. Error bars = S.E.M. (Courtesy of MF Cordeiro: The role of transforming growth factor beta in the conjunctival scarring response following glaucoma filtration surgery, PhD Thesis, University of London).

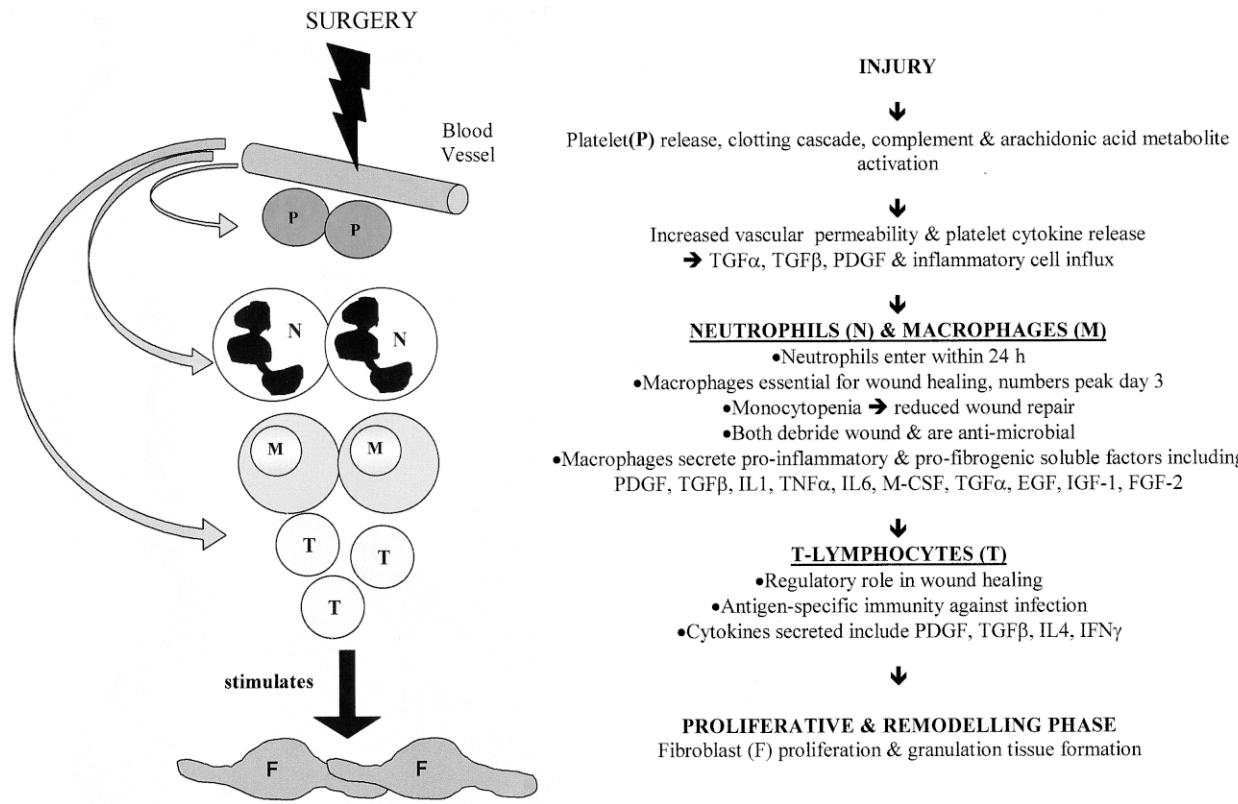


Fig. 2. The inflammatory stage of wound healing: a summary of the biochemical and cellular events in the inflammatory phase of wound healing.

is primarily active during the inflammatory stage, and this is summarized in Fig. 2.²¹

Initially, after tissue damage and blood vessel rupture, there is a release of blood cells and plasma proteins into the wound site. Hemostasis is achieved by vasoconstriction of the small blood vessels in the wounded area, which lasts for about 5–0 minutes. In addition, blood vessel rupture exposes subendothelial collagen, which stimulates platelet aggregation and activation of the intrinsic coagulation cascade, resulting in clot formation.²⁴⁷ In 1 ml of peripheral blood there are normally 2–7 million neutrophils, 1–3 million lymphocytes and 0.2–0.8 million monocytes, so it is inevitable that the bleeding accompanying the beginning of an operation will introduce a large number of inflammatory cells into the wounded area. Indeed, the importance of hemostasis during glaucoma filtration surgery to avoid promoting excessive scarring postoperatively is well recognized.

Components of the clotting cascade, in particular Hageman factor, activate the innate immune system and stimulate the generation of the vasoactive peptide bradykinin.¹³⁴

Platelets also release growth factors such as TGF α , TGF β , and PDGF, which are chemotactic and mito-

genic to inflammatory cells.^{33,142} The degree of inflammation in a wound healing response is thought to be related to the concentration of cytokines initially released by platelets. Fetal porcine platelets contain lower quantities of the cytokines TGF β and PDGF-AB, which may in part explain the reduced macrophage activation and inflammatory response associated with fetal wound repair.¹⁷⁸

The innate immune system is the body's nonspecific early defense against invading pathogens. It is comprised of the complement cascade and phagocytic populations of neutrophils and macrophages, which clear the wound site of debris and infectious agents. The complement cascade plays a major role in the early events of conjunctival wound healing by initiating inflammation through amplifying the original injury signal and by stimulating the accumulation of mitogens and chemoattractants to the injury site.

Activated complement products, such as C3a and C5a, activate the prostaglandins and other components of arachidonic acid metabolism which stimulate an increase in vascular permeability.¹⁰⁵ They are chemotactic for phagocytic cells, such as neutrophils and monocytes. The combination of all these actions results in a further influx of inflammatory cells. Subsequently, the activated arachidonic acid metabo-

lites stimulate these inflammatory cells to release other vasoactive factors, such as leukotrienes, platelet activating factor, and histamine.²⁴¹ Complement products can also coat microbes so that they are more effectively taken up by the phagocytic cells in a process known as opsonization.

B. NEUTROPHILS IN WOUND HEALING

Neutrophils are the first inflammatory cells of the immune system to enter the wound area, accumulating within 6 hours and disappearing by the third day after wounding. They are attracted to the site of injury by chemoattractants, such as C5a, platelet factor 4 (PF4), leukotrienes, TGF β , TNF α , IL1, and bacterial products.^{40,79,108,233,238} Their movement into the extracellular matrix is helped by receptors on the vascular endothelium, called selectins, and integrin receptors on the neutrophils.²²⁸

The main function of these cells is to phagocytose bacteria and foreign material in the wound, and they may be important in clearing red blood cells from the wound site.²²⁵ They may also be a source of proinflammatory cytokines such as IL1 α , IL1 β , and TNF α .¹²⁵ Although neutrophils are the first inflammatory cells to appear in wounds, they are not thought to be essential for normal wound repair, because if they are deficient and as long as an infection is not present, their function can be taken over by macrophages, and healing will occur normally.²²⁵

C. MACROPHAGES IN WOUND HEALING

The next inflammatory cells to enter the wound are monocytes, becoming the predominant inflammatory cell type at about 12 hours after injury.¹²⁹ They reach peak numbers on about the third day, but start to decrease by the fifth day. They are attracted to the wound area by chemoattractants, such as TGF β and PF4, and are activated to become macrophages by factors released by platelets and phagocytosing substances, such as fibronectin and collagen.^{32,79,246}

Unlike neutrophils, macrophages are essential for normal wound healing. This has been demonstrated by the studies of Leibovitch and Ross, who used a combination of steroids and local antimacrophage serum to induce a systemic monocytopenia and to eliminate any local tissue macrophages. This deficiency produced a significant reduction in wound debridement and fibrogenesis in guinea pig skin wounds.¹⁵¹ Also, reepithelialization was delayed, and fibroblasts did not begin to appear until the fifth day (in control wounds they were the predominant cell type by this day). They were located at the wound margins only and did not proliferate greatly.

Macrophages are important in recruiting and activating fibroblasts and other inflammatory cells. They

produce numerous soluble factors that stimulate fibroblast proliferation, and when macrophages are injected into wounds they promote wound healing.^{75,152} Moreover, corneal wound healing studies have shown that macrophages are potent stimulators of angiogenesis and collagen synthesis in a cell number dependent fashion.¹²⁷ Only one group of investigators has suggested that macrophages could act as negative regulators of fibroblast proliferation. Fukasawa et al demonstrated that the supernatant from cultured rabbit peritoneal macrophages was able to inhibit fibroblast proliferation; however, it is important to note that the fibroblasts were serum-deprived.¹⁰⁷ Therefore, in general, macrophages are thought of as stimulators of wound healing.

Macrophages produce enzymes such as collagenase and elastase, which debride the injury area and release anti-microbial factors, such as oxygen radicals and nitric oxide.^{52,158} Importantly, they provide a link between the innate and specific parts of the immune system.

D. THE REGULATORY ROLE OF LYMPHOCYTES IN WOUND HEALING

It has long been known that lymphocytes are present in healing wounds.⁸¹ Lymphocytes migrate into human skin wounds soon after the macrophages, appearing by the first day. Peak numbers develop between the eighth and fourteenth days after wounding and may persist for as long as 4 months.¹⁶⁰ In a rat wound-healing model, about 70% of the inflammatory cells were T cells by the tenth day after wounding.⁴³

Lymphocytes comprise the body's specific immune response to injury and infection. T cells become activated when they recognize antigen presented to them by macrophages, subsequently resulting in the proliferation of antigen-specific T cells. The cytokines they release can activate additional immune cells in the vicinity, including other T cells, macrophages, and polymorphs in a process called *bystander activation*.

Both CD4+ (helper/effector) and CD8+ (cytotoxic) T cell subsets were found in a rat skin wound-healing model.¹⁰⁰ They first appeared from day 5 post-wounding, peaked on day 7 and persisted until day 10. In this model, there were a greater number of lymphocytes in the deeper aspects of wounds, and the CD4+T cells outnumbered the CD8+T cells in a 2:1 ratio. Although both subsets have been identified in the conjunctiva of glaucoma patients who have failed filtration surgery at 3 months, it is not yet fully known what role the different subsets may have in the development of normal and pathological scarring.¹⁷⁴ Fig. 3 summarizes the role of T lymphocytes in wound healing.

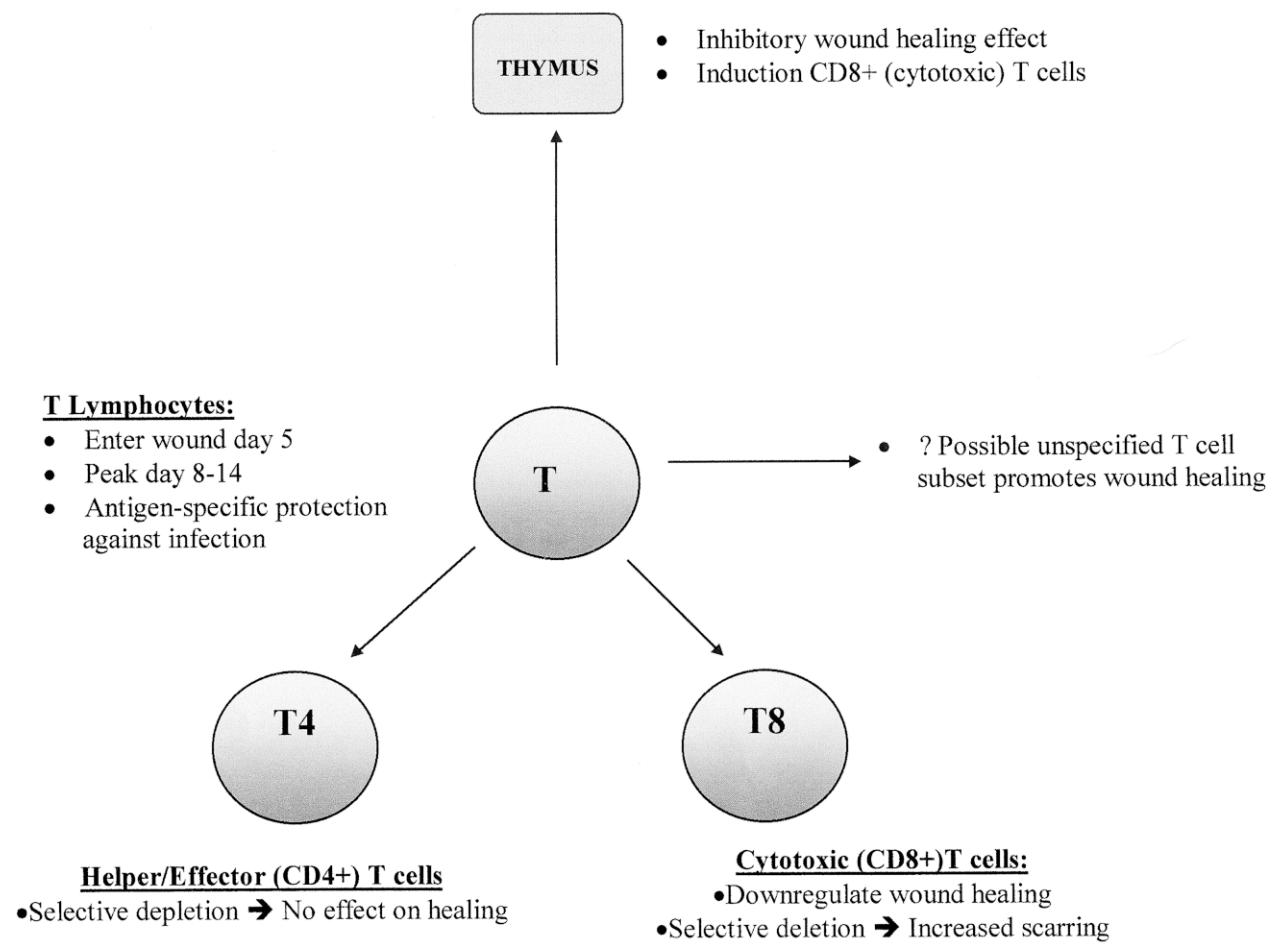


Fig. 3. The role of T lymphocytes in wound healing.

T cells are able to secrete factors that are either stimulatory or inhibitory to fibroblasts.^{237,239} For instance, Neilson et al¹⁷¹ and Postlethwaite et al¹⁹⁶ showed that, depending on culture conditions, lymphocytes produced a soluble factor, which either promoted or inhibited fibroblast collagen production. Breslin et al demonstrated that the mainly T cell infiltrate harvested from rat wounds on day 10 suppressed the growth of other T cells.⁴³ Furthermore, some researchers have found that activated T cells can inhibit fibroblast proliferation, possibly through the action of membrane-associated interferon-gamma.⁶² Finally, activated T cells may affect fibroblast migration by stimulating increased expression of the proteolytic enzyme, matrix metalloproteinase-1 (MMP-1).^{53,62} This could be important in enabling fibroblasts to contract the wound during the later stages of wound healing.

Other investigators have used experimental models that modulate lymphocyte function to demonstrate the regulatory role of lymphocytes in wound healing. Agents such as Vitamin A, arginine, and growth hormone, by stimulating T cell function, also

increase collagen deposition and wound breaking strength.^{22,86}

Conversely, the administration of agents such as steroids, retinoic acid, citral, and cyclosporin A, which all depress T cell function, are associated with impaired wound healing.⁹⁹

It is thought that the thymus probably exerts an inhibitory effect on normal wound healing, perhaps by increasing T cell regulatory activity. T cells normally mature in the thymus, leaving it to enter the blood and go to peripheral lymphoid tissues. Thymectomized rats have a deficiency in CD8+T cell induction, and this is associated with an increased wound healing response attributable to greater collagen deposition. This can be reversed when thymic grafts are implanted or when T cells are reconstituted in the originally athymic mice.^{24,25} When thymic hormones are administered, they stimulate a decrease in wound healing with a reduction in wound breaking strength and collagen deposition.²³

Depletion studies have provided further insight into the possible regulatory role of lymphocytes in wound repair.^{20,183} Firstly, global T cell depletion re-

sulted in impaired wound healing with a reduction in wound breaking strength and collagen deposition. Secondly, when the CD8+T cell subset was selectively removed, this produced a greatly increased wound healing reaction, suggesting a role for this subset in the down-regulation of wound repair. However, although simultaneous depletion of both CD4+ and CD8+T cells resulted in an increased wound healing reaction, when the CD4+T cell subset was selectively depleted, there was no change in wound healing. The authors suggested that there might exist an as yet unspecified T cell subset that could be responsible for promoting wound healing.

Recent research has focused on the existence of regulatory or anergic T cells, which may have a role in the prevention of autoimmune diseases.^{55,106} These cells can actively inhibit the immune response of other T cells. There has been no research to investigate whether they have any function in regulating wound healing.

In summary, from the research to date, we surmise that T cells play a regulatory role in wound healing. They may start off by stimulating macrophages and fibroblasts, which could then be followed by the CD8+T cells in some way switching off wound repair. Questions remain regarding how the stimulatory and inhibitory factors interact to finely tune the healing response and what events signal the immune system to switch from a stimulatory to inhibitory role. We propose that the purpose of the immune system is not only to protect the individual from an infection while a wound resolves, but also to stimulate the development and regulate progress to the next stages of the wound healing reaction.

The majority of fibroblasts are derived from local tissue fibrocytes. However, recently a novel population of bloodborne fibroblast-like cells called *fibrocytes* has been described that may play a minor role in the earliest events following wounding.^{51,60} They appear to make up about 0.5% of peripheral blood leukocytes, and enter the site of injury within 24 hours after wounding. In addition, these cells produce proinflammatory cytokines and chemokines, which may attract other inflammatory cells into the injury site. However, further research is still required to verify their possible role and contribution to wound repair.

III. Inflammatory Cytokines and Growth Factors in Wound Healing

Lymphocytes and macrophages exert many of their effects through the production of cytokines and growth factors. The research that has been conducted has used various techniques, including wound healing models to identify their presence in

vivo and to study their effects on scarring when applied artificially, and the investigation of their effects on fibroblast function in vitro. Their actions are the subject of a number of reviews.^{33,211} The following section deals with the individual cytokines researched and presents the evidence for their effects in wound healing (Table 1).

A. PLATELET-DERIVED GROWTH FACTOR (PDGF) AND TRANSFORMING GROWTH FACTOR BETA (TGF β)

Two of the most important profibrogenic cytokines are PDGF and TGF β , which are both produced by macrophages, lymphocytes, platelets, and fibroblasts, although the main producers of TGF β are macrophages and fibroblasts.^{16,17,136} Significant amounts of both TGF β and PDGF have been detected in human wound fluids, with the highest concentration of PDGF developing immediately after surgery.⁸⁵

PDGF is the most potent in vivo chemoattractant to other macrophages and fibroblasts, and it promotes increased granulation tissue formation when tested in vivo in a model of skin wound repair.^{185,188,229} Although it does not directly stimulate fibroblast collagen synthesis, it does increase deposition of glycosaminoglycans and fibronectin.^{189,190} PDGF probably exerts its effects indirectly in two ways: first, by increasing wound cellularity, and second, by inducing macrophages and fibroblasts to produce increased amounts of TGF β . This in turn stimulates increased collagen synthesis.¹⁸⁷ It also stimulates fibroblast mitogenesis, including human Tenon's fibroblast proliferation in vitro.⁹²

Mustoe et al in 1989 and Cromack et al in 1993 used models of impaired wound healing to investigate the actions of both PDGF and TGF β .^{67,169} First, total body irradiation was used to produce a monocytopenia with no concomitant effect on fibroblast numbers. This resulted in a significant decrease in wound breaking strength associated with a decrease in cellularity at the wound site. Topical PDGF wound application had no effect in improving wound breaking strength, even though it induced a local increase in fibroblast numbers; however, topical TGF β did accelerate wound healing and increased wound strength. Second, megavolt electron beam irradiation of the skin was used to reduce skin fibroblast function, sparing bone marrow function. In contrast, the application of TGF β had no effect on wound healing, but when PDGF was applied, there was an increase in wound strength with an associated increase in the number of macrophages and fibroblasts. These results suggest that macrophages are essential producers of PDGF, which mediates its wound healing effects indirectly by recruiting other

TABLE 1

The Cellular Sources and Effects of Growth Factors and Cytokines in Wound Healing

Cytokine	Cell Source	Cytokine Effects
TGF β	<ul style="list-style-type: none"> • Platelets • Macrophages • T-lymphocytes • Fibroblasts 	<ul style="list-style-type: none"> • Stimulates fibroblast migration, proliferation, and increased collagen synthesis • Chemoattractant to macrophages • Stimulates angiogenesis • Inhibits epithelial and endothelial migration and proliferation • TGF-β1, -β2, -β3 identified in mouse wound healing model
PDGF	<ul style="list-style-type: none"> • Platelets • Macrophages • T-lymphocytes • Fibroblasts • Epithelial cells • Endothelial cells • Smooth muscle cells 	<ul style="list-style-type: none"> • Chemoattractant to macrophages and fibroblasts • Stimulates fibroblast, endothelial, and epithelial cell proliferation • Stimulates glycosaminoglycans and fibronectin production • Stimulates TGFβ secretion
TGF α and EGF	<ul style="list-style-type: none"> • Platelets • Macrophages • Epithelial cells 	<ul style="list-style-type: none"> • Stimulates epithelial chemotaxis and proliferation • Stimulates angiogenesis • Stimulates fibroblast migration • Stimulates fibronectin synthesis
IGF-I	<ul style="list-style-type: none"> • Macrophages • T-lymphocytes • Epithelial cells • Endothelial cells • Fibroblasts • Smooth muscle cells 	<ul style="list-style-type: none"> • Stimulates fibroblast migration, proliferation, ECM synthesis, and contraction • Stimulates angiogenesis • Stimulates epithelial cell migration and proliferation
TNF α	<ul style="list-style-type: none"> • Macrophages • T-lymphocytes 	<ul style="list-style-type: none"> • Detected early in wound healing (first few days); probably pro-inflammatory • Synergistic with PDGF • Stimulates angiogenesis • Stimulates fibroblast secretion of M-CSF • Stimulates fibroblast proliferation in vitro • Stimulates angiogenesis • Stimulates fibroblast proliferation in vitro • Stimulates fibroblast secretion of M-CSF • May inhibit collagen production
IL 1	<ul style="list-style-type: none"> • Macrophages • T-lymphocytes 	<ul style="list-style-type: none"> • Stimulates fibroblast proliferation in vitro • Stimulates fibroblast secretion of M-CSF • May inhibit collagen production • Detected in sponges in wounds • May be involved in cell recruitment and activation • Does not affect human Tenon fibroblast proliferation • Chemotactic to fibroblasts • Stimulates collagen and fibronectin synthesis • Inhibits collagen synthesis • Stimulates fibroblast migration, proliferation, ECM synthesis, and contraction • Stimulates angiogenesis • Stimulates epithelial cell migration and ECM synthesis
IL 6	<ul style="list-style-type: none"> • Macrophages • T-lymphocytes 	
IL 4	<ul style="list-style-type: none"> • T-lymphocytes 	
Interferons bFGF	<ul style="list-style-type: none"> • T-lymphocytes • Macrophages • Endothelial cells 	

inflammatory cells and fibroblasts and inducing them to release profibrogenic cytokines such as TGF β , whereas fibroblasts are probably the main secretors of TGF β , which exerts its effects by acting directly on fibroblasts.

Our laboratory and others have found TGF β to be potently profibrogenic in conjunctival wound healing. It appears to be a potent inducer of human Tenon's fibroblast proliferation, migration, and collagen production, as well as stimulating angiogenesis and acting as a chemoattractant to other macrophages and fibroblasts.^{138,194,201,204,246} TGF β stimulates collagen production by increasing fibroblast expression of mRNA for collagen types I and III.²⁴³ In contrast to PDGF, TGF β increases granulation tissue for-

mation by selectively inducing increased maturation of collagen bundles.¹⁹⁰

All three isoforms (TGF- β 1, - β 2, - β 3) are expressed by human conjunctival fibroblasts in vitro, but only TGF- β 2 has been identified in vivo in unwounded human conjunctival stroma.^{76,180} Using a pig skin wound healing model, Levine et al showed TGF- β 2 and - β 3 staining in fibroblasts and the inflammatory infiltrate associated with the healing dermis.¹⁵⁵ Recent research by ourselves has suggested that all 3 TGF β isoforms appear to have similar actions in stimulating scarring.^{65,66} This is contrary to Shah et al and others, who showed that TGF- β 1 and - β 2 promoted scarring in rodent wound healing, whereas TGF- β 3 inhibited it.^{219,220} These somewhat

contrasting results may well be attributable to differences between species, the wound healing models used, and the anatomical site studied, although it has been suggested that TGF- β 2 may not be involved in human conjunctival scarring at all, as it is present to a similar degree in the aqueous of both nonglaucomatous and successfully filtered-glaucoma patients.⁸⁸ However, this viewpoint is somewhat in opposition with most of the literature that suggests that TGF β is a profibrogenic cytokine.²³⁴

TGF β expression is low during fetal scarless wound repair; however, if its level is artificially increased in such wounds, it provokes an inflammatory infiltrate associated with scar formation.¹⁴⁵ The addition of TGF β will accelerate the healing of several types of wounds.^{29,168,198} Its application increases the collagen content and breaking strength of normal wounds and steroid and adriamycin-impaired wounds.^{149,186} The addition of antibodies against TGF β reduces scarring after experimental glaucoma filtration surgery, and recently our laboratory has started clinical trials to test the efficacy of anti-human TGF β antibody to inhibit conjunctival scarring.^{64,218–220}

B. OTHER IMPORTANT CYTOKINES IN WOUND HEALING

Ford et al measured certain cytokine levels in implanted sponges collected from healing mouse skin wounds, and reported that significantly higher levels of macrophage-produced IL1, TNF α , IL6 and M-CSF were detected, compared to basal serum levels.¹⁰¹ However, cytokines produced by lymphocytes, such as IL2, IL3, and IL4, were not detected. The failure to detect these cytokines is surprising, but could be explained by the possible insensitivity of the tests used or perhaps because the relevant cytokines were not investigated, as, clearly, lymphocytes do in some way participate in wound healing. Although the presence of certain T cell cytokines has not been demonstrated *in vivo*, Postlethwaite et al has shown that IL4 was chemotactic to fibroblasts and stimulated increased collagen and fibronectin production *in vitro*.^{193,195}

Highest levels of mRNA for IL1 and TNF α exist at about 12–24 hours after wounding in a mouse model.¹²⁵ IL1 is an early proinflammatory cytokine, which facilitates neutrophil and monocyte entry into the wound by increasing their adhesion to vascular endothelial cells and also stimulates angiogenesis.^{40,135} In addition, IL1 may have a catabolic role during the early stages of wound healing, first, by increasing collagenase expression and second, because it has been shown to inhibit collagen synthesis when added to lattice cultures of human skin fibroblasts.¹¹² Its effects on fibroblast proliferation are less well defined; it stimulated *in vitro* human Tenon's

and dermal fibroblast proliferation, but had no effect on rat skin wound healing when administered subcutaneously.^{70,147,232}

TNF α is also a proinflammatory cytokine, and some, but not all, research suggests that it may be able to directly stimulate fibroblast activity.⁹⁴ It increases human Tenon's and diploid fibroblast proliferation, induces angiogenesis, and stimulates monocytes to secrete M-CSF.^{70,150,179,236} Like IL1, TNF α increased collagen synthesis only in serum-deprived fibroblasts. However, it increased glycosaminoglycans synthesis and collagenase production by fibroblasts cultured in both serum-deprived and serum-containing medium, suggesting a possible role for it in early wound catabolism.⁸⁴

IL6 is a multifunctional cytokine produced by macrophages and some activated T cells. It appears to have different effects, depending on the target cell; Cunliffe et al found that IL6 did not stimulate proliferation in human Tenon's fibroblasts *in vitro*.⁷⁰ It did not stimulate proliferation of fibroblasts derived from sarcoid lungs, but it did when fibroblasts derived from lungs with diffuse interstitial fibrosis were used.²²¹ M-CSF appears to regulate several macrophage functions, including phagocytic activity, superoxide production and chemotaxis.^{30,240,245}

The interferons are a heterogenous group of cytokines consisting of the Type I interferons (alpha- and beta-interferon) and Type II interferon (gamma-interferon). Alpha-interferon and beta-interferon are structurally dissimilar but are both involved in inhibiting viral infections. Gamma-interferon is produced by activated T cells and is involved in activating macrophages and promoting T cell differentiation during an immune response.

The interferons are thought to be equally effective in reducing wound healing by inhibiting collagen synthesis without affecting fibroblast proliferation.^{83,131,148,170} Gamma-interferon decreases collagen types I and III mRNA expression in fibroblasts derived from patients with the fibrosing disease scleroderma; it has successfully improved clinical parameters, such as skin score and range of limb motion, in scleroderma patients in a small prospective, non-randomized trial.^{132,203} Also, intralesional injections of alpha-interferon have reduced keloidal scarring.³⁵

There has been only one ophthalmic clinical trial to date to test the beneficial effects of intraoperative and postoperative subconjunctival injections of alpha-interferon in reducing wound healing after glaucoma filtration surgery.^{113–115} The success rate was 79% after 2 years follow-up, which was not significantly different to a success rate of 89% when 5-flourouracil was used. Alpha-IFN did not appear to offer any advantages over 5-flourouracil in terms of better intraocular pressure control and fewer side effects.

TGF α , which is produced by macrophages, stimulates epithelial cell, fibroblast, and endothelial cell mitogenesis and angiogenesis.³³ EGF, also secreted by macrophages, may stimulate fibroblast and epithelial cell chemotaxis and increase fibronectin production, which provides the initial scaffolding for clot formation during wound healing.^{3,172} TGF α appears to be a more potent angiogenic factor than EGF. IGF-I, produced by both macrophages and lymphocytes, stimulates fibroblast and endothelial cell mitogenesis.³³ Finally, FGF-2, also secreted by macrophages, stimulates fibroblast and endothelial cell mitogenesis, and both FGF-2 (basic) and VEGF stimulate angiogenesis.²²⁴

It is important to remember that the wound healing response is a dynamic process, with cytokines appearing and disappearing at different time-points. This may provide difficulties in identifying and understanding the wound healing effects of individual cytokines. Furthermore, it is also important to take into account the possible interactions of the multiple cytokines in the wound healing milieu.⁹¹ The study of their combined effects is certainly one of the challenges of future research.

IV. The Immune System in Persistent Conjunctival Scarring

A. POSSIBLE MECHANISMS OF PATHOGENESIS

Changes in the microenvironment might be able to affect the progression of wound healing, and, indeed, dysfunction of the immune response might profoundly modulate the quality and degree of wound repair. Fig. 4 represents the important balance required to ensure an appropriate wound healing response.



Fig. 4. An inflamed filtration bleb is predisposed to scar more aggressively and has a higher risk of failing filtration surgery. The cellular mechanisms that promote the development of this proinflammatory environment are only partially understood.

There is considerable evidence to suggest that abnormal inflammatory cell/fibroblast interactions, particularly those involving T lymphocytes and fibroblasts, might be involved in the pathogenesis of several fibrotic diseases and persistent conjunctival scarring. In a histological study of keloid scars, Martin et al showed that a leukocytic and fibroblast infiltrate could persist for up to several years in these lesions.¹⁶⁰ T cell/fibroblast interactions have also been implicated in fibrotic diseases, such as idiopathic pulmonary fibrosis and scleroderma. Lung T cells taken from patients with idiopathic pulmonary fibrosis stimulate human lung fibroblasts to increase their collagen production, although they also inhibit fibroblast proliferation.²¹⁶ In the bleomycin-induced model of pulmonary fibrosis, increased staining of macrophages for TGF β is associated with increased collagen production.¹³⁷ Again, in scleroderma, tissues are infiltrated with mononuclear inflammatory cells, which express increased amounts of TGF β .²⁰⁶

Hitchings and Grierson demonstrated that early trabeculectomy failures were associated with a marked inflammatory reaction and increased fibroblast numbers (Fig. 4).¹²² In contrast though, late failures look quite different histopathologically, with a bleb wall of microkeloid appearance, consisting of fibrous tissue and only occasional mononuclear inflammatory cells.² In addition, immunostaining of conjunctival biopsies taken from repeat trabeculectomy patients who have failed their first trabeculectomies at 3 months suggests that lymphocytes are increased in number and that more are activated, staining positively for IL2.¹⁷⁴ Furthermore, we know that the application of topical steroids, by downregulating the immune system and reducing the number of inflammatory cells, is associated with reduced

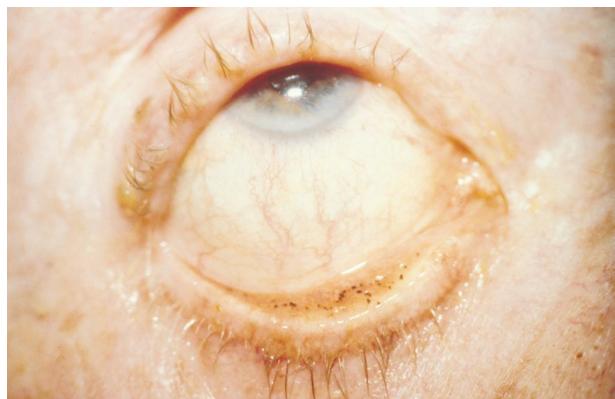


Fig. 5. This conjunctiva has developed adenochrome deposits following the use of adrenaline drops. Topical anti-glaucoma medications can adversely affect the success rate for glaucoma filtration surgery. The use of multiple medications (β -blockers, pilocarpine and sympathomimetics) reduces the surgical success rate to 45%.

TABLE 2

Evidence for the Importance of Inflammatory Cells in the Pathogenesis of Excessive Conjunctival Scarring

Evidence	Findings
Fetal wound repair	<ul style="list-style-type: none"> Normal wound healing: no inflammatory reaction and no scar tissue formation.
Depletion studies	<ul style="list-style-type: none"> Addition of TGFβ and PDGF→inflammatory infiltrate and scarring^{4,145} Systemic steroids and anti-macrophage serum produce monocytopenia→reduced wound healing^{67,127,151,152,169}
Effects of systemic drugs	<ul style="list-style-type: none"> Vitamin A, arginine, growth hormone increase T cell function and increase wound healing^{22,86,100}
Depletion studies	<ul style="list-style-type: none"> Thymectomy→decrease in induction of suppressor T cells→increased wound healing²⁵ Depletion of suppressor/cytotoxic T cell subset→increased wound healing¹⁸³
Other fibrotic conditions of body	<ul style="list-style-type: none"> Keloid lesions: increased lymphocytes and fibroblasts¹⁶⁰ Lymphocyte infiltration of tissues from patients with idiopathic pulmonary fibrosis and scleroderma, and increased expression of TGFβ^{137,216}
Fibrosing conjunctival diseases	<ul style="list-style-type: none"> OCP and DICC: Increased macrophages, increased CD4 and CD8 T cells, increased MHC II expression²⁰⁰ Increased TGFβ, PDGF, TNFα expression⁹⁰
Excessive conjunctival wound healing post filtration surgery	<ul style="list-style-type: none"> Association between early trabeculectomy failures and marked inflammatory postoperative reaction¹²² Trabeculectomy failures at 3 months contain increased CD4 and CD8 T cells¹⁷⁴ Multiple, > 3 years glaucoma drop use increases the no. macrophages and lymphocytes in conjunctiva of patients who scar aggressively post-operatively^{48,49} High risk for scarring patients (uveitic and black patients) contain increased macrophages, lymphocytes and fibroblasts⁴⁷ Topical steroids increase trabeculectomy success rates¹² May directly modulate fibroblast function²³⁵
Pharmacological modulation of inflammatory cells	

conjunctival wound healing and significantly better trabeculectomy success rates.²³⁰ Therefore, it seems reasonable to suggest that overactivity of the immune system may play an important role in the pathogenesis of at least early trabeculectomy failures. Table 2 displays the evidence for the importance of inflammatory cells in the pathogenesis of excessive conjunctival scarring.

B. PREDISPOSING FACTORS

1. Previous Topical Drug Use

What factors might stimulate the immune system to promote persistent scarring? The chronic medical treatment that most glaucoma patients take before undergoing filtration surgery may make a significant contribution by changing the cellular composition of the conjunctiva (Fig. 5).^{42,212,222} Broadway et al found that patients who had used multiple eye-drops for more than 3 years appeared to have significantly altered conjunctival cell profiles, showing an increase in the number of inflammatory cells and fibroblasts.⁴⁸ A few studies disagree with these findings; Baun et al found that the conjunctival cell profile of glaucoma patients did not change after 4 years of medical therapy, and Smith et al, using a rabbit model, found the same.^{28,227} In addition, Gwynn et al did not find any difference in the number of inflammatory cells and fibroblasts between glaucoma

patients who were better controlled after filtration surgery than those who were not so well controlled.¹¹⁹ However, Broadway's research is important because it demonstrated that these changes were associated with an increased risk of aggressive scarring and failed glaucoma filtration surgery (Fig. 6).⁴⁹

Research suggests that the irritating effects of these drops might stimulate a chronic inflammatory reaction predisposing to persistent scarring.³¹ The active compounds themselves do not appear to directly stimulate fibroblast proliferation,^{69,244} but studies have shown that the topical application of just the preservatives used in eyedrops can induce a considerable inflammatory reaction.³¹ The preservatives in glaucoma medications seem to be able to provoke quite a toxic reaction; with increased expression of the inflammatory cell marker, HLA-DR, and apoptotic markers, Fas, FasL, and APO2.7, even in the absence of overt clinical inflammation.^{26,27,44,77} Therefore, chronic use of antiglaucoma drops may somehow be able to stimulate a persistent inflammatory response in the conjunctiva of these patients, which may then predispose them to overproduce proinflammatory and profibrogenic cytokines when they are wounded. Clinically, such a process may be associated with a red eye. Such a process could ultimately affect the chances of successful glaucoma filtration surgery.

From research into the ocular fibrosing diseases,

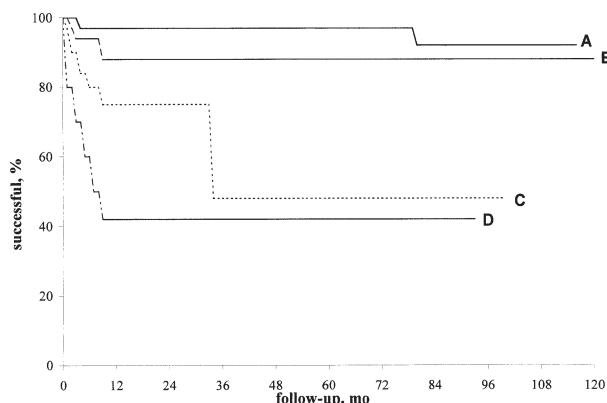


Fig. 6. Multiple topical glaucoma therapy and the success of trabeculectomy surgery. Kaplan-Meier survival analysis for glaucoma patients who underwent trabeculectomy, stratified by topical therapy regimens (Reprinted from Broadway DC et al⁴⁹ with permission of the authors and the American Medical Association [Copyright 1994, American Medical Association]). A = minimal therapy, B = β -blockers, C = β -blockers + miotic, D = β -blockers + miotic + sympathomimetic.

we have also learned a great deal about the possible role of the immune system in the pathogenesis of aggressive conjunctival scarring, such as ocular cicatricial pemphigoid and drug-induced conjunctival cicatrization.^{37,45,88} The main morphological features in ocular cicatricial pemphigoid are the infiltration of the substantia propria by inflammatory cells, followed by fibrosis of the substantia propria by new connective tissue.^{10,36,103,207} CD8+ and CD4+ T cell numbers are increased, as well as neutrophils and macrophages, with increased MHC II expression by both fibroblasts and macrophages.^{39,200} TGF β appears to be an important cytokine as its expression is increased, and greater disease activity and progression seem to be associated with increased conjunctival inflammation.^{38,89,90} The pathological changes in drug-induced conjunctival cicatrization are similar to those found in ocular cicatricial pemphigoid.^{182,197} Again, the profibrogenic cytokines implicated in this condition appear to be TGF β , PDGF, bFGF and TNF α , which, as has been mentioned before, are all produced by inflammatory cells.

2. Other High Risk Patients

There are fewer studies that examine the immune system's contribution to aggressive conjunctival scarring in other high-risk patients. Some studies, but not all, suggest that the conjunctiva of Afro-Caribbean and uveitic patients may contain an increased number of macrophages, lymphocytes, and fibroblasts.^{46,47,162} Differences in healing depending on age are largely unsubstantiated, although recent research with a murine incisional model suggests that

delayed entry of inflammatory cells and decreased expression of TGF β with age may affect the quality and degree of wound healing.¹³⁻¹⁵ The conjunctiva of younger patients does not appear to show a change in inflammatory cell counts.^{1,56,118} These studies were generally small in number and did not always look at the very young age groups. Differences in conjunctival wound healing according to age have largely been unstudied.

V. The Resolution of the Inflammatory Phase in Wound Healing

Importantly, as the wound healing response resolves, it is necessary for the immune system to deactivate itself and reduce cell numbers in order to restore immune homeostasis, since a persistent, hugely expanded population of activated inflammatory cells could be quite damaging.⁸ This, therefore, involves the death of large numbers of cells that had originally entered and proliferated at the wound site. It is thought that at the peak of the immune response, T cell death occurs through a process called activation-induced cell death or Fas-mediated apoptosis. Apoptosis is a gene-directed process whereby a cell induces its own death. Apoptotic cells have a characteristic morphology, with condensation of the nucleus and cytoplasm and nuclear fragmentation within an intact cell membrane. This form of cell death does not result in the release of the cell contents into the exterior, which might otherwise excite an inflammatory response. Fas-mediated apoptosis consists of interactions between activated T cells mediated by their surface molecules, FasL, and its receptor, Fas.^{50,80} These interactions result in a cascade of specific enzymatic reactions, which ultimately result in the programmed death of these T cells. Toward the end of the immune response, T cell numbers are reduced by nutrient or cytokine deprivation-mediated apoptosis and, although the initiating event is different, the mechanism of death is similar.⁷

Therefore, if the immune response fails to resolve properly, it is conceivable that persistently activated inflammatory cells might continue to secrete profibrogenic cytokines and promote the development of an aggressive scarring reaction. It is now known that stromal cells, including *in vitro* fibroblasts and endothelial and epithelial cells, can prevent cytokine-deprivation mediated T cell apoptosis.^{116,128}

Indeed, excessive fibroblast-mediated T cell survival seems to be a feature of chronic inflammatory conditions, such as eczema and rheumatoid arthritis.^{191,209} Recently, our group and others have identified that fibroblasts produce the soluble factor, interferon-beta, which is responsible for preventing cytokine-deprivation mediated T cell death.¹⁵⁹ It has also been suggested that TGF β may prevent Fas-

mediated T cell apoptosis.^{54,109} It also seems that intraocular levels of TGF β can be correlated with the amount of scarring induced in fibrotic diseases like proliferative vitreoretinopathy.⁶³ We could therefore speculate that in persistent conjunctival scarring, an abnormal cycle of interactions between T cells and fibroblasts might be set up, with the fibroblasts keeping the T lymphocytes alive, which, in turn, stimulate the fibroblasts to continue to produce scar tissue. In terms of aggressive conjunctival wound healing, this is an area of research requiring further evaluation.

VI. The Role of the Immune System in Other Ocular Fibrosing Diseases

Although this review concentrates on the impact of the immune system on conjunctival wound healing, it is important to note that interactions between inflammatory cells and fibroblasts may play a role in other ocular inflammatory or wound healing disorders. The pathophysiology of proliferative vitreoretinopathy can be compared to a wound healing reaction in a specialized tissue. Several studies have shown that inflammatory cells e.g., macrophages and lymphocytes together with fibroblastic-like retinal pigment epithelial (RPE) cells, make up the periretinal membranes.^{57,133} Some studies have suggested that the macrophages seem to be of the acute inflammatory subtype when the intraocular proliferation is more severe and that the lymphocytes present in proliferative vitreoretinopathy tissue are activated and belong to both subsets.^{57,58,93}

It is likely that the growth factors secreted by these inflammatory cells contribute to the pathogenesis of PVR; IL1, IL6, TNF α , IFN γ , TGF β , and PDGF have all been identified in samples of PVR vitreous or membranes.¹⁵⁷ Limb et al also found that there was significant HLA-DR expression in proliferative vitreoretinopathy membranes and suggested that local autoimmune responses may be occurring within them.¹⁵⁶ The body of evidence strongly suggests that key interactions between inflammatory cells and fibroblastic-like RPE cells contribute significantly to the pathogenesis of the disease.

The chronicity of the autoimmune disorder thyroid eye disease may also be caused by fibroblast/T cell interactions. Although the nature of the autoantigen initiating the disease is not well characterized, it is known that the associated inflammatory infiltrate releases a combination of cytokines, which stimulate orbital fibroblasts to produce glycosaminoglycans.^{18,120} It would be interesting to examine whether orbital fibroblast-mediated prevention of T cell apoptosis might contribute to the chronic inflammatory process of thyroid eye disease. Finally, recent evidence suggests that macrophages partici-

pate in corneal wound healing after excimer laser keratotomy.¹⁷⁵ It has been suggested that their ability to influence the development of corneal haze should be examined.

VII. The Modulation of the Immune System in Wound Healing

A. CURRENT THERAPIES

The success rate of glaucoma filtration surgery may be optimized through the use of a combination of pharmacological agents that modulate the inflammatory and proliferative phases of the wound healing reaction.^{86,215,248} The most commonly clinically used drugs are corticosteroids and the antimetabolites, 5-fluorouracil (5-FU) and mitomycin-C (MMC), which improve trabeculectomy success rates in both low and high risk for scarring patients.^{11,59} Topical antiprostaglandins, which are known inhibitors of inflammation, have not proved to be useful in improving the outcome of fistulizing surgery, although newer agents have not been tested.¹⁶³

Trabeculectomy surgery is more successful with post-operative steroids, and systemic steroids do not appear to have an advantage over topical administration.^{12,202} Other studies have examined the effect of steroids on reducing bleb inflammation and fibrosis.^{110,167}

Corticosteroids have potent antiinflammatory and immunoregulatory effects.⁷¹ The mechanisms of their effects are still not entirely understood. When taken systemically, corticosteroids cause a redistribution of circulating peripheral blood lymphocytes to the bone marrow, resulting in a lymphocytopenia and monocytopenia.⁹⁵ Steroids can augment suppressor T cell activity and inhibit T cell proliferation and antigen presentation.²¹³ Monocytes may be especially sensitive to steroids, with suppression of bactericidal activity. They also appear to inhibit the access of neutrophils and monocytes to the inflammatory site.⁹⁶ Steroids reduce vascular permeability and may decrease the secretion of proinflammatory cytokines. Dexamethasone-treated full-thickness skin wounds in mice showed significantly reduced expression of mRNA for IL1 α , IL1 β and TNF α .^{125,210} Therefore, topically applied steroids probably exert most of their effects by decreasing the inflammatory phase of conjunctival wound healing, by reducing the influx of inflammatory cells and by decreasing the production of pro-fibrogenic cytokines.

Some authors have suggested that steroids may affect fibroblast function directly.^{19,82,235} They have been shown to inhibit the in vitro contraction of collagen gels, and they appear to have a biphasic effect on fibroblast proliferation, inhibiting proliferation at low doses and stimulating it at higher doses.^{41,74} Clinically, steroid use has been reportedly associated

with a change in the morphology of filtration blebs, causing thin, cystic bleb walls and even necrosis.^{111,167,205,231} Miller et al found that topical steroids had only a temporary delaying effect on fibroblast proliferation in a rabbit model of glaucoma fistulizing surgery. The main effects appeared to be caused by a reduction in the number of inflammatory cells and a decrease in aqueous chamber flare, reflecting the stabilization of the blood-aqueous barrier.¹⁶⁵ These somewhat different findings may be because the rabbit displays more aggressive scarring, and, therefore, further research into the effects of steroids on human conjunctival wound healing would be of interest.

Mitomycin-C and 5-FU are chemotherapeutic agents, which exert their effects on malignant cells by inducing cell death or apoptosis. Mitomycin-C acts on cells by damaging their DNA, through cross-linking bases in the same or adjacent DNA strands. 5-FU acts on proliferating cells primarily by inhibiting thymidylate synthetase and preventing the synthesis of DNA. With respect to inhibiting conjunctival scarring, our laboratory has shown that these agents induce human Tenon's fibroblast apoptosis, as well as inhibiting fibroblast proliferation, migration, and collagen contraction.^{34,68,139,140,176} Weinreb suggested that 5-FU could have a beneficial effect on inflammation; he reported that some of his patients with inflammatory glaucomas required less steroid after surgery than before.²⁴² Although these drugs are primarily used in trabeculectomy surgery because of their actions on fibroblast function, they almost certainly exert the same effects on inflammatory cells, as shown by their virtual absence in biopsies taken from Mitomycin-C-treated blebs.^{181,223}

The use of these drugs is not entirely ideal, because they do have potential side effects. Corticosteroids may be associated with cataractogenesis, raised intraocular pressure, and infection. Hypotony, endophthalmitis, and toxic effects on the corneal epithelium are complications associated with antimetabolite use. Furthermore, it is not completely understood yet why some patients at high risk for scarring still fail filtration surgery, despite the use of these drugs.^{61,226} One reason may be because growth-arrested human Tenon's fibroblasts, when tested in vitro, are still able to migrate and secrete certain growth factors, such as TGF β . This may allow some patients to overcome the antiscarring effects of the antimetabolites.¹⁷⁷

B. FUTURE THERAPIES

The antimetabolites and steroids have certainly had a significant impact on improving the success rate of filtration surgery. However, future treatments could consist of more targeted therapy that would be directed at blocking specific cytokines or inter-

ing with important inflammatory cell/fibroblast interactions that might lead to persistent scarring.

As mentioned before, this approach is already being investigated. Gillies et al found that postoperative subconjunctival injections of alpha-interferon compared similarly to 5-FU in terms of reducing wound healing and controlling intraocular pressure after 2 years of follow-up. Unfortunately, combination therapy did not appear to have an increased effect.¹¹⁴ Research in our laboratory is currently in progress to investigate the effects of antihuman TGF β antibody in reducing conjunctival wound healing.⁶⁴

Cyclosporin is a powerful immunosuppressive agent most commonly used in preventing organ transplant rejection. In ophthalmology it may be of clinical benefit in the treatment of T cell-mediated chronic inflammatory conditions such as vernal and atopic keratoconjunctivitis.^{123,214} It exerts its effects by blocking the proliferation and activation of T cells by inhibiting IL2 transcription and the expression of the receptor for IL2.^{104,144} In atopic keratoconjunctivitis, it reduces the inflammatory infiltrate, normalizes the CD4:CD8 ratio, and reduces IL2, gamma-IFN, and HLA-DR expression.¹²¹ It may also be able to modulate wound healing, and encouraging data have been reported in a rabbit wound healing model.^{74,173,184} However, it appears to have no effect on corneal epithelial and rat skin wound healing, and it may stimulate TGF β and induce gingival overgrowth.^{6,87,98,143,192} Therefore, the role of cyclosporin in modulating excessive conjunctival scarring needs to be evaluated further. It could be useful in modulating the inflammatory cell profile of high-risk patients preoperatively or to reduce the inflammatory condition of those patients with severe chronic inflammatory conjunctivitis preoperatively. Alternatively, it could be an important steroid sparing agent for those patients at high-risk for scarring who are intolerant of intensive topical steroids.

Some recent research has highlighted the potential role of mast cells in wound healing.¹⁵⁴ They are present in hypertrophic scars, and one of the secretory products, histamine, may be involved in promoting in vitro wound closure.^{141,146,153} Disodium cromoglycate has been shown to reduce skin wound healing and collagen synthesis in a rat wound healing model.⁷³ Another secretory product, heparin, has been shown to be necessary for stabilizing other secretory granules inside the mast cells.^{102,126} In addition, it appears to inhibit human corneal fibroblast proliferation in vitro.⁷⁸ Recently, we have shown that the interoperative combination of 5FU and heparin significantly reduces the risk of postoperative retinal scarring in patients undergoing retinal detachment surgery, and it may decrease the risk of the development of proliferative retinopathy (Chang L et al: *Ophthalmology*, in press).

Another immunomodulatory approach to wound healing might be the use of cytokines, which could play some role in the development of ocular immune privilege. This phenomenon involves a numbers of factors that protect the interior of the eye from the damaging inflammatory effects accompanying immune responses. One novel cytokine that has been implicated is IL10.^{72,97}

In conclusion, many of the constituents of the immune system participate in the wound healing response. It is evident that a fine balance in the immune system's response to wounding is required to ensure the correct degree of repair. The challenge will be to develop specific therapies to switch wound healing on or off depending on the clinical response of the patient, to avoid some of the unsatisfactory side-effects of the drugs currently used, and to ultimately improve the surgical success rate of glaucoma filtration surgery.

Method of Literature Search

A comprehensive international literature search was achieved through the use of Medline, Winspurs, and PubMed—databases for basic science and clinical medical research—as well as obtaining articles cited by authors but not found on these databases. Search words included: *wound healing, conjunctiva, immune system, T-lymphocyte, macrophage, glaucoma, trabeculectomy*. These were used in combinations to ensure that an exhaustive selection of the relevant references was obtained. The databases dated from 1960–1999.

Searches on the databases dated from 1960. Older references were obtained from checking older articles.

References

- Abdel-Khalek LM, Williamson J, Lee WR: Morphological changes in the human conjunctival epithelium. I. In the normal elderly population. *Br J Ophthalmol* 62: 792–9, 1978
- Addicks EM, Quigley HA, Green WR, Robin AL: Histologic characteristics of filtering blebs in glaucomatous eyes. *Arch Ophthalmol* 101: 795–8, 1983
- Adelmann-Grill BC, Wach F, Cully Z, et al: Chemotactic migration of normal dermal fibroblasts towards epidermal growth factor and its modulation by platelet-derived growth factor and transforming growth factor-beta. *Eur J Cell Biol* 51: 322–6, 1990
- Adolph VR, DiSanto SK, Bleacher JC, et al: The potential role of the lymphocyte in fetal wound healing. *J Pediatr Surg* 28: 1316–20, 1993
- Adzick NS, Harrison MR, Glick PL, et al: Comparison of fetal, newborn, and adult wound healing by histologic, enzyme-histochemical, and hydroxyproline determinations. *J Pediatr Surg* 20: 315–9, 1985
- Ahuja SS, Shrivastav S, Danielpour D, et al: Regulation of transforming growth factor-beta 1 and its receptor by cyclosporine in human T lymphocytes. *Transplantation* 60: 718–23, 1995
- Akbar AN, Borthwick N, Salmon M, et al: The significance of low bcl-2 expression by CD45RO T cells in normal individuals and patients with acute viral infections. The role of apoptosis in T cell memory. *J Exp Med* 178: 427–38, 1993
- Akbar AN, Salmon M: Cellular environments and apoptosis: tissue microenvironments control activated T-cell death. *Immunology Today* 18: 72–6, 1997
- Allansmith MR, Greiner JV, Baird RS: Number of inflammatory cells in the normal conjunctiva. *Am J Ophthalmol* 86: 250–9, 1978
- Andersen SR, Jensen OA, Kristensen EB, Norn MS: Benign mucous membrane pemphigoid. 3. Biopsy. *Acta Ophthalmol* 52: 455–63, 1974
- Anonymous: Five-year follow-up of the Fluorouracil Filtering Surgery Study. The Fluorouracil Filtering Surgery Study Group. *Am J Ophthalmol* 121: 349–66, 1996
- Araujo SV, Spaeth GL, Roth SM, Starita RJ: A ten-year follow-up on a prospective, randomized trial of postoperative corticosteroids after trabeculectomy. *Ophthalmology* 102: 1753–9, 1995
- Ashcroft GS, Herrick SE, Tarnuzzer RW, et al: Human ageing impairs injury-induced *in vivo* expression of tissue inhibitor of matrix metalloproteinases (TIMP)-1 and -2 proteins and mRNA. *J Pathol* 183: 169–76, 1997
- Ashcroft GS, Horan MA, Ferguson MW: Aging is associated with reduced deposition of specific extracellular matrix components, an upregulation of angiogenesis, and an altered inflammatory response in a murine incisional wound healing model. *J Invest Dermatol* 108: 430–7, 1997
- Ashcroft GS, Horan MA, Ferguson MW: Aging alters the inflammatory and endothelial cell adhesion molecule profiles during human cutaneous wound healing. *Lab Invest* 78: 47–58, 1998
- Assoian RK, Fleurdeley BE, Stevenson HC, et al: Expression and secretion of type beta transforming growth factor by activated human macrophages. *Proc Natl Acad Sci USA* 84: 6020–4, 1987
- Assoian RK, Komoriya A, Meyers CA, et al: Transforming growth factor-beta in human platelets. Identification of a major storage site, purification, and characterization. *J Biol Chem* 258: 7155–60, 1983
- Bahn RS, Heufelder A: Pathogenesis of Graves' ophthalmopathy. *N Engl J Med* 329: 1468–75, 1993
- Ball SF, Vinh T, Gebhardt BM: Biphasic effect of corticosteroids on protein and DNA synthesis by human subconjunctival fibroblasts in culture [abstract]. *Invest Ophthalmol Vis Sci (Suppl)* 28: S378, 1987
- Barbul A, Breslin RJ, Woodard JP, et al: The effect of *in vivo* T helper and T suppressor lymphocyte depletion on wound healing. *Ann Surg* 209: 479–83, 1989
- Barbul A, Regan MC: Immune involvement in wound healing. *Otolaryngol Clin N Am* 28: 955–68, 1995
- Barbul A, Rettura G, Levenson SM, Seifter E: Arginine: a thymotropic and wound-healing promoting agent. *Surg Forum* 28: 101–3, 1977
- Barbul A, Shawe T, Frankel HL, et al: Inhibition of wound repair by thymic hormones. *Surgery* 106: 373–6, 1989
- Barbul A, Shawe T, Rotter SM, et al: Wound healing in nude mice: a study on the regulatory role of lymphocytes in fibroplasia. *Surgery* 105: 764–9, 1989
- Barbul A, Sisto D, Rettura G, et al: Thymic inhibition of wound healing: abrogation by adult thymectomy. *J Surg Res* 32: 338–42, 1982
- Baudouin C, Garcher C, Haouat N, et al: Expression of inflammatory membrane markers by conjunctival cells in chronically treated patients with glaucoma. *Ophthalmology* 101: 454–60, 1994
- Baudouin C, Pisella PJ, Fillacier K, et al: Ocular surface inflammatory changes induced by topical antiglaucoma drugs: human and animal studies. *Ophthalmology* 106: 556–63, 1999
- Baun O, Heegaard S, Kessing SV, Prause JU: The morphology of conjunctiva after long-term topical anti-glaucoma treatment. A quantitative analysis. *Acta Ophthalmol Scand* 73: 242–5, 1995
- Beck LS, DeGuzman L, Lee WP, et al: One systemic administration of transforming growth factor-beta 1 reverses age- or glucocorticoid-impaired wound healing. *J Clin Invest* 92: 2841–9, 1993
- Becker S, Warren MK, Haskill S: Colony-stimulating factor-induced monocyte survival and differentiation into mac-

rophages in serum-free cultures. *J Immunol* 139: 3703-9, 1987

31. Becquet F, Goldschild M, Moldovan MS, et al: Histopathological effects of topical ophthalmic preservatives on rat corneoconjunctival surface. *Curr Eye Res* 17: 419-25, 1998
32. Beezhold DH, Personius C: Fibronectin fragments stimulate tumor necrosis factor secretion by human monocytes. *J Leukocyte Biol* 51: 59-64, 1992
33. Bennett NT, Schultz GS: Growth factors and wound healing: biochemical properties of growth factors and their receptors. *Am J Surg* 165: 728-37, 1993
34. Bergstrom TJ, Wilkinson WS, Skuta GL, et al: The effects of subconjunctival mitomycin-C on glaucoma filtration surgery in rabbits. *Arch Ophthalmol* 109: 1725-30, 1991
35. Berman B, Duncan MR: Short-term keloid treatment in vivo with human interferon alfa-2b results in a selective and persistent normalization of keloidal fibroblast collagen, glycosaminoglycan, and collagenase production in vitro. *J Am Acad Dermatol* 21: 694-702, 1989
36. Bernauer W, Broadway DC, Wright P: Chronic progressive conjunctival cicatrization. *Eye* 7: 371-8, 1993
37. Bernauer W, Itin PH, Kirtschig G: Cicatricial pemphigoid. *Dev Ophthalmol* 28: 46-63, 1997
38. Bernauer W, Wright P, Dart JK, et al: Cytokines in the conjunctiva of acute and chronic mucous membrane pemphigoid: an immunohistochemical analysis. *Graefes Arch Clin Exp Ophthalmol* 231: 563-70, 1993
39. Bernauer W, Wright P, Dart JK, et al: The conjunctiva in acute and chronic mucous membrane pemphigoid. An immunohistochemical analysis. The value of biopsies in the evaluation of chronic progressive conjunctival cicatrization. *Ophthalmology* 100: 339-46, 1993
40. Bevilacqua MP, Pober JS, Wheeler ME, et al: Interleukin 1 acts on cultured human vascular endothelium to increase the adhesion of polymorphonuclear leukocytes, monocytes, and related leukocyte cell lines. *J Clin Invest* 76: 2003-11, 1985
41. Blumenkranz MS, Claflin A, Hajek AS: Selection of therapeutic agents for intraocular proliferative disease. Cell culture evaluation. *Arch Ophthalmol* 102: 598-604, 1984
42. Brandt JD, Wittpenn JR, Katz LJ, et al: Conjunctival impression cytology in patients with glaucoma using long-term topical medication. *Am J Ophthalmol* 112: 297-301, 1991
43. Breslin RJ, Wasserkrug HL, Efron G, Barbul A: Suppressor cell generation during normal wound healing. *J Surg Res* 44: 321-5, 1988
44. Brignole F, De Saint-Jean M, Goldschild M, et al: Expression of Fas-Fas ligand antigens and apoptotic marker APO2.7 by the human conjunctival epithelium. Positive correlation with class II HLA DR expression in inflammatory ocular surface disorders. *Exp Eye Res* 67: 687-97, 1998
45. Broadway D: Drug-induced conjunctival cicatrization. *Develop Ophthalmol* 28: 86-101, 1997
46. Broadway D, Grierson I, Hitchings R: Racial differences in the results of glaucoma filtration surgery: are racial differences in the conjunctival cell profile important? *Br J Ophthalmol* 78: 466-75, 1994
47. Broadway DC, Bates AK, Lightman SL, et al: The importance of cellular changes in the conjunctiva of patients with uveitic glaucoma undergoing trabeculectomy. *Eye* 7: 495-501, 1993
48. Broadway DC, Grierson I, O'Brien C, Hitchings RA: Adverse effects of topical antiglaucoma medication. I. The conjunctival cell profile. *Arch Ophthalmol* 112: 1437-45, 1994
49. Broadway DC, Grierson I, O'Brien C, Hitchings RA: Adverse effects of topical antiglaucoma medication. II. The outcome of filtration surgery. *Arch Ophthalmol* 112: 1446-54, 1994
50. Brunner T, Mogil RJ, LaFace D, et al: Cell-autonomous Fas (CD95)/Fas-ligand interaction mediates activation-induced apoptosis in T-cell hybridomas. *Nature* 373: 441-4, 1995
51. Bucala R, Spiegel LA, Chesney J, et al: Circulating fibrocytes define a new leukocyte subpopulation that mediates tissue repair. *Molec Med* 1: 71-81, 1994
52. Buchmuller-Rouiller Y, Mauel J: Macrophage activation for intracellular killing as induced by calcium ionophore. Correlation with biologic and biochemical events. *J Immunol* 146: 217-23, 1991
53. Burger D, Rezzonico R, Li JM, et al: Imbalance between interstitial collagenase and tissue inhibitor of metalloproteinases 1 in synoviocytes and fibroblasts upon direct contact with stimulated T lymphocytes: involvement of membrane-associated cytokines. *Arthritis Rheum* 41: 1748-59, 1998
54. Cerwenka A, Kovar H, Majdic O, Holter W: Fas- and activation-induced apoptosis are reduced in human T cells pre-activated in the presence of TGF-beta 1. *J Immunol* 156: 459-64, 1996
55. Chai JG, Bartok I, Chandler P, et al: Anergic T cells act as suppressor cells in vitro and in vivo. *Eur J Immunol* 29: 686-92, 1999
56. Chan CC, Nussenblatt RB, Ni M, et al: Immunohistochemical markers in the normal human epibulbar conjunctiva from fetus to adult. *Arch Ophthalmol* 106: 215-7, 1988
57. Charteris DG, Hiscott P, Grierson I, Lightman SL: Lymphocytes in epiretinal membranes. *Ophthalmology* 99: 1364-7, 1992
58. Charteris DG, Hiscott P, Grierson I, Lightman SL: Inflammatory cells in proliferative vitreoretinopathy subretinal membranes. *Ophthalmology* 100: 43-6, 1993
59. Chen CW, Huang HT, Bair JS, Lee CC: Trabeculectomy with simultaneous topical application of mitomycin-C in refractory glaucoma. *J Ocul Pharmacol* 6: 175-82, 1990
60. Chesney J, Metz C, Stavitsky AB, et al: Regulated production of type I collagen and inflammatory cytokines by peripheral blood fibrocytes. *J Immunol* 160: 419-25, 1998
61. Cheung JC, Wright MM, Murali S, Pederson JE: Intermediate-term outcome of variable dose mitomycin C filtering surgery. *Ophthalmology* 104: 143-9, 1997
62. Chizzolini C, Rezzonico R, Ribbens C, et al: Inhibition of type I collagen production by dermal fibroblasts upon contact with activated T cells: different sensitivity to inhibition between systemic sclerosis and control fibroblasts. *Arthritis Rheum* 41: 2039-47, 1998
63. Connor TBJ, Roberts AB, Sporn MB, et al: Correlation of fibrosis and transforming growth factor-beta type 2 levels in the eye. *J Clin Invest* 83: 1661-6, 1989
64. Cordeiro MF, Gay JA, Khaw PT: Human anti-transforming growth factor- β 2 antibody: a new glaucoma anti-scarring agent. *Invest Ophthalmol Vis Sci* 40: 2225-34, 1999
65. Cordeiro MF, Reichel MB, Gay JA, et al: Transforming growth factor- β 1, - β 2, and - β 3 in vivo: effects on normal and mitomycin C-modulated conjunctival scarring. *Invest Ophthalmol Vis Sci* 40: 1975-82, 1999
66. Cox DA: Transforming growth factor-beta 3. *Cell Biol Int* 19: 357-71, 1995
67. Cromack DT, Porras-Reyes B, Purdy JA, et al: Acceleration of tissue repair by transforming growth factor beta 1: identification of in vivo mechanism of action with radiotherapy-induced specific healing deficits. *Surgery* 113: 36-42, 1993
68. Crowston JG, Akbar AN, Constable PH, et al: Antimetabolite-induced apoptosis in Tenon's capsule fibroblasts. *Invest Ophthalmol Vis Sci* 39: 449-54, 1998
69. Cunliffe I, McIntyre C, Rees R, Rennie I: The effect of topical beta-blocker medications on the proliferation and viability of human Tenon's capsule fibroblasts in tissue culture. *German J Ophthalmol* 4: 167-74, 1995
70. Cunliffe IA, Richardson PS, Rees RC, Rennie IG: Effect of TNF, IL-1, and IL-6 on the proliferation of human Tenon's capsule fibroblasts in tissue culture. *Br J Ophthalmol* 79: 590-5, 1995
71. Cupps TR, Fauci AS: Corticosteroid-mediated immunoregulation in man. *Immunol Rev* 65: 133-55, 1982
72. D'Orazio TJ, Niederkorn JY: A novel role for TGF- β and IL-10 in the induction of immune privilege. *J Immunol* 160: 2089-98, 1998
73. Dabrowski R, Drobniak J: The effect of disodium cromoglycate on the skin wound healing and collagen content in the wounds of rats. *Acta Physiologica Polonica* 41: 195-8, 1990

74. Damji KF, Rootman J, Palcic B, Thurston G: Pharmacological modulation of human subconjunctival fibroblast behaviour in vitro. *Ophthalmic Surg* 21: 31–43, 1990

75. Danon D, Kowatch MA, Roth GS: Promotion of wound repair in old mice by local injection of macrophages. *Proc Natl Acad Sci USA* 86: 2018–20, 1989

76. De-Quan L, Lee S-B, Tseng SCG: Differential expression and regulation of TGF- β 1, TGF- β 2, TGF- β 3, TGF- β RI, TGF- β RII, and TGF- β RIII in cultured human corneal, limbal and conjunctival fibroblasts. *Curr Eye Res* 19: 154–61, 1999

77. De Saint JM, Brignole F, Bringuier AF, et al: Effects of benzalkonium chloride on growth and survival of Chang conjunctival cells. *Invest Ophthalmol Vis Sci* 40: 619–30, 1999

78. Denk PO, Knorr M: Effect of heparin on human corneal fibroblast proliferation in vitro with and without growth factor stimulation. *Graefes Arch Clin Exp Ophthalmol* 237: 342–7, 1999

79. Deuel TF, Senior RM, Chang D, et al: Platelet factor 4 is chemotactic for neutrophils and monocytes. *Proc Natl Acad Sci USA* 78: 4584–7, 1981

80. Dhein J, Walczak H, Baumler C, et al: Autocrine T-cell suicide mediated by APO-1/(Fas/CD95). *Nature* 373: 438–41, 1995

81. Diegelmann RF, Kim JC, Lindblad WJ, et al: Collection of leukocytes, fibroblasts, and collagen within an implantable reservoir tube during tissue repair. *J Leukocyte Biol* 42: 667–72, 1987

82. Duke-Elder S, Ashton N: Action of cortisone on tissue reactions of inflammation and repair with special reference to the eye. *Br J Ophthalmol* 35: 695–707, 1951

83. Duncan MR, Berman B: Gamma interferon is the lymphokine and beta interferon the monokine responsible for inhibition of fibroblast collagen production and late but not early fibroblast proliferation. *J Exp Med* 162: 516–27, 1985

84. Duncan MR, Berman B: Differential regulation of collagen, glycosaminoglycan, fibronectin, and collagenase activity production in cultured human adult dermal fibroblasts by interleukin 1-alpha and beta and tumor necrosis factor-alpha and beta. *J Invest Dermatol* 92: 699–706, 1989

85. Dvorch VM, Murphey RJ, Matsuoka J, Grotendorst GR: Changes in growth factor levels in human wound fluid. *Surgery* 112: 18–23, 1992

86. Ehrlich HP, Hunt TK: Effects of cortisone and vitamin A on wound healing. *Ann Surg* 167: 324–8, 1968

87. Eisinger DR, Sheil AG: A comparison of the effects of cyclosporin A and standard agents on primary wound healing in the rat. *Surg Gynecol Obstet* 160: 135–8, 1999

88. Elder MJ: The role of cytokines in chronic progressive conjunctival cicatrization. *Develop Ophthalmol* 28: 159–75, 1997

89. Elder MJ, Bernauer W, Leonard J, Dart JK: Progression of disease in ocular cicatricial pemphigoid. *Br J Ophthalmol* 80: 292–6, 1996

90. Elder MJ, Dart JK, Lightman S: Conjunctival fibrosis in ocular cicatricial pemphigoid—the role of cytokines. *Exp Eye Res* 65: 165–76, 1997

91. Elias JA, Freundlich B, Kern JA, Rosenblom J: Cytokine networks in the regulation of inflammation and fibrosis in the lung. *Chest* 97: 1439–45, 1990

92. Ellis DG, Cheng Q, Lee DA: The effects of growth factors on Tenon's capsule fibroblasts in serum-free culture. *Curr Eye Res* 15: 27–35, 1996

93. Esser P, Heimann K, Wiedemann P: Macrophages in proliferative vitreoretinopathy and proliferative diabetic retinopathy: differentiation of subpopulations. *Br J Ophthalmol* 77: 731–3, 1993

94. Fahey TJ, Sherry B, Tracey KJ, et al: Cytokine production in a model of wound healing: the appearance of MIP-1, MIP-2, cachectin/TNF and IL-1. *Cytokine* 2: 92–9, 1990

95. Fauci AS: Mechanisms of corticosteroid action on lymphocyte subpopulations. I. Redistribution of circulating T and B lymphocytes to the bone marrow. *Immunology* 28: 669–80, 1975

96. Fauci AS, Dale DC, Balow JE: Glucocorticosteroid therapy: mechanisms of action and clinical considerations. *Ann Intern Med* 84: 304–15, 1976

97. Ferguson TA, Griffith TS: A vision of cell death: insights into immune privilege. *Immunol Rev* 156: 167–84, 1997

98. Filipcic M, Phan TM, Zhoa T-Z, et al: Topical cyclosporine A and corneal wound healing. *Cornea* 11: 546–52, 1992

99. Fishel R, Barbul A, Wasserkrug HL, et al: Cyclosporine A impairs wound healing in rats. *J Surg Res* 34: 572–5, 1983

100. Fishel RS, Barbul A, Beschorner WE, et al: Lymphocyte participation in wound healing. Morphologic assessment using monoclonal antibodies. *Ann Surg* 206: 25–9, 1987

101. Ford HR, Hoffman RA, Wing EJ, et al: Characterization of wound cytokines in the sponge matrix model. *Arch Surg* 124: 1422–8, 1989

102. Forsberg E, Pejler G, Ringvall M, et al: Abnormal mast cells in mice deficient in a heparin-synthesizing enzyme. *Nature* 400: 773–6, 1999

103. Foster CS: Cicatricial pemphigoid. *Trans Am Ophthalmol Soc* 84: 527–663, 1986

104. Foxwell BM, Simon J, Herrero JJ, et al: Anti-CD3 antibody-induced expression of both p55 and p75 chains of the high affinity interleukin-2 receptor on human T lymphocytes is inhibited by cyclosporin A. *Immunology* 69: 104–9, 1990

105. Frank MM, Fries LF: The role of complement in inflammation and phagocytosis. *Immunol Today* 12: 322–6, 1991

106. Frasca L, Carmichael P, Lechler R, Lombardi G: Anergic T cells effect linked suppression. *Eur J Immunol* 27: 3191–7, 1997

107. Fukasawa M, Bryant SM, Nakamura RM, diZerega GS: Modulation of fibroblast proliferation by postsurgical macrophages. *J Surg Res* 43: 513–20, 1987

108. Gamble JR, Harlan JM, Klebanoff SJ, Vadas MA: Stimulation of the adherence of neutrophils to umbilical vein endothelium by human recombinant tumor necrosis factor. *Proc Natl Acad Sci USA* 82: 8667–71, 1985

109. Genestier L, Kasibhatla S, Brunner T, Green DR: Transforming growth factor beta1 inhibits Fas ligand expression and subsequent activation-induced cell death in T cells via downregulation of c-Myc. *J Exp Med* 189: 231–9, 1999

110. Giangiaco J, Dueker DK, Adelstein E: The effect of pre-operative subconjunctival triamcinolone administration on glaucoma filtration. I. Trabeculectomy following subconjunctival triamcinolone. *Arch Ophthalmol* 104: 838–41, 1986

111. Giangiaco J, Dueker DK, Adelstein EH: Histopathology of triamcinolone in the subconjunctiva. *Ophthalmology* 94: 149–53, 1987

112. Gillery P, Couston F, Pujol JP, Borel JP: Inhibition of collagen synthesis by interleukin-1 in three-dimensional collagen lattice cultures of fibroblasts. *Experientia* 45: 98–101, 1989

113. Gillies M, Su T, Sarossy M, Hollows F: Interferon-alpha 2b inhibits proliferation of human Tenon's capsule fibroblasts. *Graefes Arch Clin Exp Ophthalmol* 231: 119–21, 1993

114. Gillies MC, Brooks AM, Young S, et al: A randomized phase II trial of interferon-alpha2b versus 5-fluorouracil after trabeculectomy. *Aust NZ J Ophthalmol* 27: 37–44, 1999

115. Gillies MC, Goldberg I, Young SH, Su T: Glaucoma filtering surgery with interferon- α -2b. *J Glaucoma* 2: 229–35, 1993

116. Gombert W, Borthwick NJ, Wallace DL, et al: Fibroblasts prevent apoptosis of IL-2-deprived T cells without inducing proliferation: a selective effect on Bcl-XL expression. *Immunology* 89: 397–404, 1996

117. Goss AN: Intra-uterine healing of fetal rat oral mucosal, skin and cartilage wounds. *J Oral Pathol* 6: 35–43, 1977

118. Gwynn DR, Stewart WC, Hennis HL, et al: The influence of age upon inflammatory cell counts and structure of conjunctiva in chronic open-angle glaucoma. *Acta Ophthalmologica* 71: 691–5, 1993

119. Gwynn DR, Stewart WC, Pitts RA, et al: Conjunctival structure and cell counts and the results of filtering surgery. *Am J Ophthalmol* 116: 464–8, 1993

120. Heufelder A, Bahn R: Modulation of Graves' orbital fibroblast proliferation by cytokines and glucocorticoid receptor agonists. *Invest Ophthalmol Vis Sci* 35: 120-7, 1994

121. Hingorani M, Calder VL, Buckley RJ, Lightman S: The immunomodulatory effect of topical cyclosporin in atopic keratoconjunctivitis. *Invest Ophthalmol Vis Sci* 40: 392-9, 1999

122. Hitchings RA, Grierson I: Clinicopathological correlation in eyes with failed fistulizing surgery. *Trans Ophthalmol Soc UK* 103: 84-8, 1983

123. Holland EJ, Olsen TW, Ketcham JM, et al: Topical cyclosporin A in the treatment of anterior segment inflammatory disease. *Cornea* 12: 413-9, 1993

124. Hopkinson-Woolley J, Hughes D, Gordon S, Martin P: Macrophage recruitment during limb development and wound healing in the embryonic and fetal mouse. *J Cell Sci* 107: 1159-67, 1994

125. Hubner G, Brauchle M, Smola H, et al: Differential regulation of pro-inflammatory cytokines during wound healing in normal and glucocorticoid-treated mice. *Cytokine* 8: 548-56, 1996

126. Humphries DE, Wong GW, Friend DS, et al: Heparin is essential for the storage of specific granule proteases in mast cells. *Nature* 400: 769-72, 1999

127. Hunt TK, Knighton DR, Thakral KK, et al: Studies on inflammation and wound healing: angiogenesis and collagen synthesis stimulated in vivo by resident and activated wound macrophages. *Surgery* 96: 48-54, 1984

128. Hyde H, Borthwick NJ, Janossy G, et al: Upregulation of intracellular glutathione by fibroblast-derived factor(s): enhanced survival of activated T cells in the presence of low Bcl-2. *Blood* 89: 2453-60, 1997

129. Issekutz TB, Issekutz AC, Movat HZ: The in vivo quantitation and kinetics of monocyte migration into acute inflammatory tissue. *Am J Pathol* 103: 47-55, 1981

130. Jay JL, Allan D: The benefit of early trabeculectomy versus conventional management in primary open angle glaucoma relative to severity of disease. *Eye* 3: 528-35, 1989

131. Jimenez SA, Freundlich B, Rosenbloom J: Selective inhibition of human diploid fibroblast collagen synthesis by interferons. *J Clin Invest* 74: 1112-6, 1984

132. Kahan A, Amor B, Menkes CJ, Strauch G: Recombinant interferon-gamma in the treatment of systemic sclerosis. *Am J Med* 87: 273-7, 1989

133. Kampik A, Kenyon KR, Michels RG, et al: Epiretinal and vitreous membranes. *Arch Ophthalmol* 99: 1445-54, 1981

134. Kaplan AP: Hageman factor-dependent pathways: mechanism of initiation and bradykinin formation. *Federation Proc* 42: 3123-7, 1983

135. Kaushansky K, Lin N, Adamson JW: Interleukin 1 stimulates fibroblasts to synthesize granulocyte-macrophage and granulocyte colony-stimulating factors. Mechanism for the hematopoietic response to inflammation. *J Clin Invest* 81: 92-7, 1988

136. Kehrl JH, Wakefield LM, Roberts AB, et al: Production of transforming growth factor beta by human T lymphocytes and its potential role in the regulation of T cell growth. *J Exp Med* 163: 1037-50, 1986

137. Khalil N, Berezny O, Sporn M, Greenberg AH: Macrophage production of transforming growth factor beta and fibroblast collagen synthesis in chronic pulmonary inflammation. *J Exp Med* 170: 727-37, 1989

138. Khaw PT, Occleston NL, Larkin G, et al: The effects of growth factors on human ocular fibroblast proliferation, migration and collagen production [abstract]. *Invest Ophthalmol Vis Sci* 35 (Suppl):1898, 1994

139. Khaw PT, Sherwood MB, MacKay SL, et al: Five-minute treatments with fluorouracil, floxuridine, and mitomycin have long-term effects on human Tenon's capsule fibroblasts. *Arch Ophthalmol* 110: 1150-4, 1992

140. Khaw PT, Ward S, Porter A, et al: The long-term effects of 5-fluorouracil and sodium butyrate on human Tenon's fibroblasts. *Invest Ophthalmol Vis Sci* 33: 2043-52, 1992

141. Kischer CW, Bunce H 3rd, Shetlah MR: Mast cell analyses in hypertrophic scars, hypertrophic scars treated with pressure and mature scars. *J Invest Dermatol* 70: 355-7, 1978

142. Knighton DR, Ciresi K, Fiegel VD, et al: Stimulation of repair in chronic, nonhealing, cutaneous ulcers using platelet-derived wound healing formula. *Surg Gynecol Obstet* 170: 56-60, 1990

143. Kossendrup D, Wiederholt M, Hoffman F: Influence of cyclosporin A, dexamethasone, and benzalkonium chloride (BAK) on corneal epithelial wound healing in the rabbit and guinea pig eye. *Cornea* 4: 177-81, 1985

144. Kronke M, Leonard WJ, Depper JM, et al: Cyclosporin A inhibits T-cell growth factor gene expression at the level of mRNA transcription. *Proc Natl Acad Sci USA* 81: 5214-18, 1984

145. Krummel TM, Michna BA, Thomas BL, et al: Transforming growth factor beta (TGF-beta) induces fibrosis in a fetal wound model. *J Pediatric Surg* 23: 647-52, 1988

146. Kupietzky A, Levi-Schaffer F: The role of mast cell-derived histamine in the closure of an in vitro wound. *Inflammation Res* 45: 176-80, 1996

147. Laato M, Heino J: Interleukin 1 modulates collagen accumulation by rat granulation tissue cells both in vivo and in vitro. *Experientia* 44: 32-4, 1988

148. Latina MA, Belmonte SJ, Park C, Crean E: Gamma-interferon effects on human fibroblasts from Tenon's capsule. *Invest Ophthalmol Vis Sci* 32: 2806-15, 1991

149. Lawrence WT, Norton JA, Sporn MB, et al: The reversal of an Adriamycin induced healing impairment with chemoattractants and growth factors. *Ann Surg* 203: 142-7, 1986

150. Leibovich SJ, Polverini PJ, Shepard HM, et al: Macrophage-induced angiogenesis is mediated by tumour necrosis factor-alpha. *Nature* 329: 630-2, 1987

151. Leibovich SJ, Ross R: The role of the macrophage in wound repair. A study with hydrocortisone and antimacrophage serum. *Am J Pathol* 78: 71-100, 1975

152. Leibovich SJ, Ross R: A macrophage-dependent factor that stimulates the proliferation of fibroblasts in vitro. *Am J Pathol* 84: 501-14, 1976

153. Leonardi A, Radice M, Fregonia IA, et al: Histamine effects on conjunctival fibroblasts from patients with vernal conjunctivitis. *Exp Eye Res* 68: 739-46, 1999

154. Levi-Schaffer F, Kupietzky A: Mast cells enhance migration and proliferation of fibroblasts into an in vitro wound. *Exp Cell Res* 188: 42-9, 1990

155. Levine JH, Moses HL, Gold LI, Nanney LB: Spatial and temporal patterns of immunoreactive transforming growth factor beta 1, beta 2, and beta 3 during excisional wound repair. *Am J Pathol* 143: 368-80, 1993

156. Limb GA, Franks WA, Munasinghe KR, et al: Proliferative vitreoretinopathy: an examination of the involvement of lymphocytes, adhesion molecules and HLA-DR antigens. *Graefes Arch Clin Exp Ophthalmol* 231: 331-6, 1993

157. Limb GA, Little BC, Meager A, et al: Cytokines in proliferative vitreoretinopathy. *Eye* 5: 686-93, 1991

158. Malawista SE, Montgomery RR, van BG: Evidence for reactive nitrogen intermediates in killing of staphylococci by human neutrophil cytoplasm. A new microbicidal pathway for polymorphonuclear leukocytes. *J Clin Invest* 90: 631-6, 1992

159. Marrack P, Kappler J, Mitchell T: Type I Interferons Keep Activated T cells Alive. *J Exp Med* 189: 521-9, 1999

160. Martin CW, Muir IF: The role of lymphocytes in wound healing. *Br J Plastic Surg* 43: 655-62, 1990

161. Martin P: Wound healing-aiming for perfect skin regeneration. *Science* 276: 75-81, 1997

162. McMillan TA, Stewart WC, Hennis HL, et al: Histologic differences in the conjunctiva of black and white glaucoma patients. *Ophthalmic Surg* 23: 762-5, 1992

163. Migdal C, Hitchings R: The developing bleb: effect of topical antiprostaglandins on the outcome of glaucoma fistulising surgery. *Br J Ophthalmol* 67: 655-60, 1983

164. Migdal C, Hitchings R: Control of chronic simple glaucoma with primary medical, surgical and laser treatment. *Trans Ophthalmol Soc UK* 105: 653-6, 1986

165. Miller MH, Grierson I, Unger WG, Hitchings RA: The effect of topical dexamethasone and preoperative beta irradiation on a model of glaucoma fistulizing surgery in the rabbit. *Ophthalmic Surg* 21: 44–54, 1990

166. Miller MH, Grierson I, Unger WI, Hitchings RA: Wound healing in an animal model of glaucoma fistulizing surgery in the rabbit. *Ophthalmic Surg* 20: 350–7, 1989

167. Molteno AC, Straughan JL, Ancker E: Control of bleb fibrosis after glaucoma surgery by anti-inflammatory agents. *S Afric Med J* 50: 881–5, 1976

168. Mustoe TA, Pierce GF, Thomason A, et al: Accelerated healing of incisional wounds in rats induced by transforming growth factor-beta. *Science* 237: 1333–6, 1987

169. Mustoe TA, Purdy J, Gramates P, et al: Reversal of impaired wound healing in irradiated rats by platelet-derived growth factor-BB. *Am J Surg* 158: 345–50, 1989

170. Nguyen KD, Hoang AT, Lee DA: Transcriptional control of human Tenon's capsule fibroblast collagen synthesis in vitro by gamma-interferon. *Invest Ophthalmol Vis Sci* 35: 3064–70, 1994

171. Nielson EG, Phillips SM, Jimenez S: Lymphokine modulation of fibroblast proliferation. *J Immunol* 128: 1484–6, 1982

172. Nishida T, Tanaka H, Nakagawa S, et al: Fibronectin synthesis by the rabbit cornea: effects of mouse epidermal growth factor and cyclic AMP analogs. *Jpn J Ophthalmol* 28: 196–202, 1984

173. Nuzzi R, Cerruti A, Finazzo C: Cyclosporine C: a study of wound-healing modulation after trabeculectomy in rabbit. *Acta Ophthalmol Scand Suppl*:48–9, 1998

174. Nuzzi R, Vercelli A, Finazzo C, Cracco C: Conjunctiva and subconjunctival tissue in primary open-angle glaucoma after long-term topical treatment: an immunohistochemical and ultrastructural study. *Graefes Arch Clin Exp Ophthalmol* 233: 154–62, 1995

175. O'Brien TP, Li Q, Ashrafi F, et al: Inflammatory Response in the Early Stages of Wound Healing after Excimer Laser Keratectomy. *Arch Ophthalmol* 116: 1470–4, 1998

176. Occleston NL, Alexander RA, Mazure A, et al: Effects of single exposures to antiproliferative agents on ocular fibroblast-mediated collagen contraction. *Invest Ophthalmol Vis Sci* 35: 3681–90, 1994

177. Occleston NL, Daniels JT, Tarnuzzer RW, et al: Single exposures to antiproliferatives: long-term effects on ocular fibroblast wound-healing behaviour. *Invest Ophthalmol Vis Sci* 38: 1998–2007, 1997

178. Olutoye OO, Yager DR, Cohen IK, Diegelmann RF: Lower cytokine release by fetal porcine platelets: a possible explanation for reduced inflammation after fetal wounding. *J Pediatric Surg* 31: 91–5, 1996

179. Oster W, Lindemann A, Horn S, et al: Tumor necrosis factor (TNF)-alpha but not TNF-beta induces secretion of colony stimulating factor for macrophages (CSF-1) by human monocytes. *Blood* 70: 1700–3, 1987

180. Pasquale LR, Dorman-Pease ME, Lutty GA, et al: Immunolocalization of TGF-beta 1, TGF-beta 2, and TGF-beta 3 in the anterior segment of the human eye. *Invest Ophthalmol Vis Sci* 34: 23–30, 1993

181. Pasquale LR, Thibault D, Dorman-Pease ME, et al: Effect of topical mitomycin C on glaucoma filtration surgery in monkeys. *Ophthalmology* 99: 14–8, 1992

182. Patten JT, Cavanagh HD, Allansmith MR: Induced ocular pseudopemphigoid. *Am J Ophthalmol* 82: 272–6, 1976

183. Peterson JM, Barbul A, Breslin RJ, et al: Significance of T-lymphocytes in wound healing. *Surgery* 102: 300–5, 1987

184. Petri JB, Schurk S, Gebauer S, Haustein UF: Cyclosporine A delays wound healing and apoptosis and suppresses activin beta-A expression in rats. *Eur J Dermatol* 8: 104–13, 1998

185. Pierce GF, Brown D, Mustoe TA: Quantitative analysis of inflammatory cell influx, procollagen type I synthesis, and collagen cross-linking in incisional wounds: influence of PDGF-BB and TGF-beta 1 therapy. *J Lab Clin Med* 117: 373–82, 1991

186. Pierce GF, Mustoe TA, Lingelbach J, et al: Transforming growth factor beta reverses the glucocorticoid-induced wound-healing deficit in rats: possible regulation in macrophages by platelet-derived growth factor. *Proc Natl Acad Sci USA* 86: 2229–33, 1989

187. Pierce GF, Mustoe TA, Lingelbach J, et al: Platelet-derived growth factor and transforming growth factor-beta enhance tissue repair activities by unique mechanisms. *J Cell Biol* 109: 429–40, 1989

188. Pierce GF, Mustoe TA, Senior RM, et al: In vivo incisional wound healing augmented by platelet-derived growth factor and recombinant c-sis gene homodimeric proteins. *J Exp Med* 167: 974–87, 1988

189. Pierce GF, Tarpley JE, Yanagihara D, et al: Platelet-derived growth factor (BB homodimer), transforming growth factor-beta 1, and basic fibroblast growth factor in dermal wound healing. *Neovessel and matrix formation and cessation of repair*. *Am J Pathol* 140: 1375–88, 1992

190. Pierce GF, Vande BJ, Rudolph R, et al: Platelet-derived growth factor-BB and transforming growth factor beta 1 selectively modulate glycosaminoglycans, collagen, and myofibroblasts in excisional wounds. *Am J Pathol* 138: 629–46, 1991

191. Pilling D, Akbar AN, Girdlestone J, et al: Interferon-beta mediates stromal cell rescue of T cells from apoptosis. *Eur J Immunol* 29: 1041–50, 1999

192. Plemons JM, Dill RE, Rees TD, et al: PDGF-B producing cells and PDGF-B gene expression in normal gingival and cyclosporin-induced overgrowth. *J Periodontol* 67: 264–70, 1996

193. Postlethwaite AE, Holness MA, Katai H, Raghaw R: Human fibroblasts synthesize elevated levels of extracellular matrix proteins in response to interleukin 4. *J Clin Invest* 90: 1479–85, 1992

194. Postlethwaite AE, Keski-Oja J, Moses HL, Kang AH: Stimulation of the chemotactic migration of human fibroblasts by transforming growth factor beta. *J Exp Med* 165: 251–6, 1987

195. Postlethwaite AE, Seyer JM: Fibroblast chemotaxis induction by human recombinant interleukin-4. Identification by synthetic peptide analysis of two chemotactic domains residing in amino acid sequences 70–88 and 89–122. *J Clin Invest* 87: 2147–52, 1991

196. Postlethwaite AE, Smith GN, Mainardi CL, et al: Lymphocyte modulation of fibroblast function in vitro: stimulation and inhibition of collagen production by different effector molecules. *J Immunol* 132: 2470–7, 1984

197. Pouliquen Y, Patey A, Foster CS, et al: Drug-induced cicatricial pemphigoid affecting the conjunctiva. Light and electron microscopic features. *Ophthalmology* 93: 775–83, 1986

198. Quaglino DJ, Nanney LB, Ditesheim JA, Davidson JM: Transforming growth factor-beta stimulates wound healing and modulates extracellular matrix gene expression in pig skin: incisional wound model. *J Invest Dermatol* 97: 34–42, 1991

199. Reichel MB, Cordeiro MF, Alexander RA, et al: New model of conjunctival scarring in the mouse eye. *Br J Ophthalmol* 82: 1072–7, 1998

200. Rice BA, Foster CS: Immunopathology of cicatricial pemphigoid affecting the conjunctiva. *Ophthalmology* 97: 1476–83, 1990

201. Roberts AB, Sporn MB, Assoian RK, et al: Transforming growth factor type beta: rapid induction of fibrosis and angiogenesis in vivo and stimulation of collagen formation in vitro. *Proc Natl Acad Sci USA* 83: 4167–71, 1986

202. Robinson DI, Lertsumitkul S, Billson FA, Robinson LP: Long-term intraocular pressure control by trabeculectomy: a ten-year life table. *Aust NZ J Ophthalmol* 21: 79–85, 1993

203. Rosenbloom J, Feldman G, Freundlich B, Jimenez SA: Inhibition of excessive scleroderma fibroblast collagen production by recombinant gamma-interferon. Association with a coordinate decrease in types I and III procollagen messenger RNA levels. *Arthritis Rheum* 29: 851–6, 1986

204. Rossi P, Karsenty G, Roberts AB et al: A nuclear factor 1 binding site mediates the transcriptional activation of a type I collagen promoter by transforming growth factor-beta. *Cell* 52: 405-14, 1988

205. Roth SM, Spaeth GL, Starita RJ, et al: The effects of postoperative corticosteroids on trabeculectomy and the clinical course of glaucoma: five-year follow-up study. *Ophthalmic Surg* 22: 724-9, 1991

206. Roumm AD, Whiteside TL, Medsger TAJ, Rodnan GP: Lymphocytes in the skin of patients with progressive systemic sclerosis. Quantification, subtyping, and clinical correlations. *Arthritis Rheum* 27: 645-53, 1984

207. Sacks EH, Jakobiec FA, Wieczorek R, et al: Immunophenotypic analysis of the inflammatory infiltrate in ocular cicatricial pemphigoid. Further evidence for a T cell-mediated disease. *Ophthalmology* 96: 236-43, 1989

208. Sacks EH, Wieczorek R, Jakobiec FA, Knowles DM: Lymphocytic subpopulations in the normal human conjunctiva. A monoclonal antibody study. *Ophthalmology* 93: 1276-83, 1986

209. Salmon M, Scheel-Toellner D, Huissoon AP et al: Inhibition of T cell apoptosis in the rheumatoid synovium. *J Clin Invest* 99: 439-46, 1997

210. Schmidt J, Fleissner S, Heimann-Weitschat I, et al: Effect of corticosteroids, cyclosporin A, and methotrexate on cytokine release from monocytes and T-cell subsets. *Immunopharmacology* 27: 173-9, 1994

211. Schultz G, Khaw PT, Oxford K et al: Growth factors and ocular wound healing. *Eye* 8: 184-7, 1994

212. Schwab IR, Linberg JV, Gioia VM, et al: Foreshortening of the inferior conjunctival fornix associated with chronic glaucoma medications. *Ophthalmology* 99: 197-202, 1992

213. Scudeletti M, Pende D, Barabino A, et al: Effect of single oral doses of prednisone and deflazacort on human lymphocyte distribution and functions. Analysis with monoclonal antibodies. *Adv Exp Med Biol* 171: 335-44, 1984

214. Secchi AG, Tognon MS, Leonardi A: Topical use of cyclosporine in the treatment of vernal keratoconjunctivitis. *Am J Ophthalmol* 110: 641-645, 1990

215. Seetner A, Morin JD: Healing of trabeculectomies in rabbits. *Can J Ophthalmol* 14: 121-5, 1979

216. Selman M, Gonzalez G, Bravo M, et al: Effect of lung T lymphocytes on fibroblasts in idiopathic pulmonary fibrosis and extrinsic allergic alveolitis. *Thorax* 45: 451-5, 1990

217. Serini G, Gabbiani G: Modulation of alpha-smooth muscle actin expression in fibroblasts by TGF-beta. *Wound Repair Regeneration* 4: 278, 1996

218. Shah M, Foreman DM, Ferguson MW: Control of scarring in adult wounds by neutralising antibody to transforming growth factor beta. *Lancet* 339: 213-4, 1992

219. Shah M, Foreman DM, Ferguson MW: Neutralising antibody to TGF-beta 1,2 reduces cutaneous scarring in adult rodents. *J Cell Sci* 107: 1137-57, 1994

220. Shah M, Foreman DM, Ferguson MW: Neutralisation of TGF-beta 1 and TGF-beta 2 or exogenous addition of TGF-beta 3 to cutaneous rat wounds reduces scarring. *J Cell Sci* 108: 985-1002, 1995

221. Shahar I, Fireman E, Topilsky M et al: Effect of IL-6 on alveolar fibroblast proliferation in interstitial lung diseases. *Clin Immunol Immunopathol* 79: 244-51, 1996

222. Sherwood MB, Grierson I, Millar L, Hitchings RA: Long-term morphologic effects of antiglaucoma drugs on the conjunctiva and Tenon's capsule in glaucomatous patients. *Ophthalmology* 96: 327-35, 1989

223. Shields MB, Scroggs MW, Sloop CM, Simmons RB: Clinical and histopathologic observations concerning hypotony after trabeculectomy with adjunctive mitomycin C. *Am J Ophthalmol* 116: 673-83, 1993

224. Shipley GD, Keeble WW, Hendrickson JE, et al: Growth of normal human keratinocytes and fibroblasts in serum-free medium is stimulated by acidic and basic fibroblast growth factor. *J Cell Physiol* 138: 511-8, 1989

225. Simpson DM, Ross R: The neutrophilic leukocyte in wound repair a study with antineutrophil serum. *J Clin Invest* 51: 2009-23, 1972

226. Skuta GL, Beeson CC, Higginbotham EJ, et al: Intraoperative mitomycin versus postoperative 5-fluorouracil in high-risk glaucoma filtering surgery. *Ophthalmology* 99: 438-44, 1992

227. Smith DL, Skuta GL, Kincaid MC, et al: The effects of glaucoma medications on Tenon's capsule and conjunctiva in the rabbit. *Ophthalmic Surg* 22: 336-40, 1991

228. Springer TA: Traffic signals for lymphocyte recirculation and leukocyte emigration: the multistep paradigm. *Cell* 76: 301-14, 1994

229. Sprugel KH, McPherson JM, Clowes AW, Ross R: Effects of growth factors in vivo. I. Cell ingrowth into porous subcutaneous chambers. *Am J Pathol* 129: 601-13, 1987

230. Starita RJ, Fellman RL, Spaeth GL, et al: Short- and long-term effects of postoperative corticosteroids on trabeculectomy. *Ophthalmology* 92: 938-46, 1985

231. Sugar HS: Clinical effect of corticosteroids on conjunctival filtering blebs; a case report. *Am J Ophthalmol* 59: 854-60, 1965

232. Thornton SC, Por SB, Walsh BJ, et al: Interaction of immune and connective tissue cells: I. The effect of lymphokines and monokines on fibroblast growth. *J Leukocyte Biol* 47: 312-20, 1990

233. Tonnesen MG, Smedly LA, Henson PM: Neutrophil-endothelial cell interactions. Modulation of neutrophil adhesiveness induced by complement fragments C5a and C5a des arg and formyl-methionyl-leucyl-phenylalanine in vitro. *J Clin Invest* 74: 1581-92, 1984

234. Tripathi RC, Li J, Chan WF, Tripathi BJ: Aqueous humor in glaucomatous eyes contains an increased level of TGF-beta 2. *Exp Eye Res* 59: 723-7, 1994

235. Van Story-Lewis PE, Tenenbaum HC: Glucocorticoid inhibition of fibroblast contraction of collagen gels. *Biochem Pharmacol* 35: 1283-6, 1986

236. Vilcek J, Palombella VJ, Henriksen-DeStefano D, et al: Fibroblast growth enhancing activity of tumor necrosis factor and its relationship to other polypeptide growth factors. *J Exp Med* 163: 632-43, 1986

237. Wahl SM, Gately CL: Modulation of fibroblast growth by a lymphokine of human T cell continuous T cell line origin. *J Immunol* 130: 1226-30, 1983

238. Wahl SM, Hunt DA, Wakefield LM et al: Transforming growth factor type beta induces monocyte chemotaxis and growth factor production. *Proc Natl Acad Sci USA* 84: 5788-92, 1987

239. Wahl SM, Wahl LM, McCarthy JB: Lymphocyte-mediated activation of fibroblast proliferation and collagen production. *J Immunol* 121: 942-6, 1978

240. Wang JM, Griffin JD, Rambaldi A, et al: Induction of monocyte migration by recombinant macrophage colony-stimulating factor. *J Immunol* 141: 575-9, 1988

241. Wedmore CV, Williams TJ: PAF, a sceretary product of polymorphonuclear leukocytes, increases vascular permeability in rabbit skin. *Br J Pharmacol* 74: 916-7, 1981

242. Weinreb RN: Adjusting the dose of 5-fluorouracil after filtration surgery to minimize side effects. *Ophthalmology* 94: 564-70, 1987

243. Westergren-Thorsson G, Antonsson P, Malmstrom A, et al: The synthesis of a family of structurally related proteoglycans in fibroblasts is differently regulated by TFG-beta. *Matrix* 11: 177-83, 1991

244. Williams DE, Nguyen KD, Shapourifar-Tehrani S, et al: Effects of timolol, betaxolol, and levobunolol on human tenon's fibroblasts in tissue culture. *Invest Ophthalmol Vis Sci* 33: 2233-41, 1992

245. Wing EJ, Ampel NM, Waheed A, Shadduck RK: Macrophage colony-stimulating factor (M-CSF) enhances the capacity of murine macrophages to secrete oxygen reduction products. *J Immunol* 135: 2052-6, 1985

246. Wiseman DM, Polverini PJ, Kamp DW, Leibovich SJ: Transforming growth factor-beta (TGF beta) is chemotactic for human monocytes and induces their expression of angio-

genic activity. *Biochem Biophys Res Comm* 157: 793–800, 1988

247. Witte MB, Barbul A: General principles of wound healing. *Surg Clin N Am* 77: 509–28, 1997

248. Woods AC: Clinical and experimental observation on the use of ACTH and cortisone in ocular inflammatory disease. *Am J Ophthalmol* 33: 1325–49, 1950

Outline

- I. Wound healing repair models
 - A. Animal models
 - B. Fetal wound repair
- II. The inflammatory phase of wound healing
 - A. Activation of the immune system in wound healing
 - B. The role of neutrophils in wound healing
 - C. The role of macrophages in wound healing
 - D. The regulatory role of lymphocytes in wound healing
- III. Inflammatory cytokines and growth factors in wound healing
 - A. Platelet-derived growth factor and transforming growth factor-beta
 - B. Other important cytokines in wound healing
- IV. The immune system in persistent conjunctival scarring

- A. Possible mechanisms of pathogenesis
- B. Predisposing factors
 - 1. Topical drug use
 - 2. Other high risk patients
- V. The resolution of the inflammatory phase in wound healing
- VI. The role of the immune system in other ocular fibrosing diseases
- VII. The modulation of the immune system in wound healing
 - A. Current therapies
 - B. Future therapies

This work was supported by Wellcome Trust Grant Nos. 055183 (LC), 048474 (MFC), 045202 (JGC), and the Medical Research Council (G9330070). It was undertaken by authors who are funded in part by the NHS Executive; the views expressed in this publication are those of the authors and not necessarily of the NHS Executive. This work was also supported by the Glaucoma Unit, Moorfields Eye Hospital NHS Trust, London, U.K.

The authors have no proprietary or commercial interest in any product or idea discussed in this article.

Reprint address: L Chang, FRCOphth, Wound Healing Research Unit, Department of Pathology, Institute of Ophthalmology, Bath Street, London, EC1V 9EL, United Kingdom